

# Bioeroding\_sponge\_assemblag es-----.pdf

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# Bioeroding sponge assemblages: the importance of substrate availability and sediment

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Despite global deterioration of coral reef health, not all reef-associated organisms are in decline. Bioeroding sponges are thought to be largely resistant to the factors that stress and kill corals, and are increasing in abundance on many reefs. However, there is a paucity of information on how environmental factors influence spatial variation in the distribution of these sponges, and how they might be affected by different stressors. We aimed to identify the factors that explained differences in bioeroding sponge abundance and assemblage composition, and to determine whether bioeroding sponges benefit from the same environmental conditions that can contribute towards coral mortality. Abundance surveys were conducted in the Wakatobi region of Indonesia on reefs characterized by different biotic and abiotic conditions. Bioeroding sponges occupied an average of 8.9% of available dead substrate and variation in abundance and assemblage composition was primarily attributed to differences in the availability of dead substrate. Our results imply that if dead substrate availability increases as a consequence of coral mortality, bioeroding sponge abundance is also likely to increase. However, bioeroding sponge abundance was lowest on a sedimented reef, despite abundant dead substrate. This suggests that not all forms of coral mortality will benefit all bioeroding sponge species, and sediment-degraded reefs are likely to be dominated by a few resilient bioeroding sponge species. Overall, we demonstrate the importance of understanding the drivers of bioeroding sponge abundance and assemblage composition in order to predict possible impacts of different stressors on reefs communities.

**Keywords:** Distribution, reef degradation, run-off, sedimentation, substrate availability

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## INTRODUCTION

Despite their ecological and economic importance, coral reefs worldwide are increasingly threatened by anthropogenic activities. Over the past 50 years, coral reefs have suffered from substantial declines in coral cover, habitat complexity and biodiversity (e.g. Bruno & Selig, 2007; Burke *et al.*, 2011). In some locations, coral cover has already been reduced by 50% or more since the 1980s (Bruno & Selig, 2007; De'ath *et al.*, 2012). Declines in spatially competitive scleractinian corals can provide populations of less competitive benthic taxa with an opportunity to expand, and there are increasing reports of substantial reef community shifts (e.g. McManus & Polsenberg, 2004; Norström *et al.*, 2009). While the majority of the literature has focused on shifts from coral to macroalgal-dominated communities, many other groups of benthic taxa can potentially benefit from coral declines (Mumby, 2009; Norström *et al.*, 2009; Cheal *et al.*, 2010; Bell *et al.*, 2013). Bioeroding sponges are of such group, and are reportedly becoming more abundant on some degraded reefs in the Caribbean (e.g. Rützler, 2002;

López-Victoria & Zea, 2004), the eastern Pacific (Carballo *et al.*, 2013), and on the Great Barrier Reef (GBR) (Schönberg & Ortiz, 2009). For example bioeroding sponges occupy up to 46% of sampled substrate on bleached reefs in the Mexican Pacific (Carballo *et al.*, 2013) and increased in abundance by 150% on the Great Barrier Reef (GBR) after two major bleaching events (Schönberg & Ortiz, 2009). Many bioeroding sponge species appear to be resilient to some of the stressors that are detrimental to corals (reviewed in Schönberg *et al.*, 2017b), and some species are aggressive spatial competitors (e.g. López-Victoria & Zea, 2005; Chaves-Fonnegra & Zea, 2007; González-Rivero *et al.*, 2017).

A wide range of organisms contribute to bioerosion (e.g. Glynn, 1997; Wisshak & Tapanila, 2008; Schönberg *et al.*, 2017b), and infestation levels of different boring sponges can vary depending on environmental conditions (e.g. Tribollet *et al.*, 2002; Tribollet & Golubic, 2005). However, sponges regularly generate the majority of internal reef bioerosion and often represent 60–90% of macroborer activity (e.g. Risk *et al.*, 1995; Mallela & Perry, 2007). Rates of erosion can be very high, with some species capable of removing over 20 kg of calcareous substrate per m<sup>2</sup> sponge surface area a year (Schönberg, 2002; Calcinaï *et al.*, 2008). Furthermore, under certain environmental conditions that favour high abundances of bioeroding sponges, and in combination with reduced coral cover/calcification, the balance

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between calcification and reef erosion could potentially be altered by sponge bioerosion, resulting in net carbonate erosion (e.g. Nava & Carballo, 2008; Perry *et al.*, 2008; Kennedy *et al.*, 2013). Moreover, bioerosion has been identified as a principal driver of the loss of reef structural complexity (e.g. Alvarez-Filip *et al.*, 2011). This is important, as three-dimensional reef structure provides critical habitat, maintaining densities and biomass of reef organisms, and is central to ecosystem services that support tourism and shoreline protection (e.g. Enochs & Manzello, 2012; Graham & Nash, 2012).

Given the potentially negative sequences of increases in bioeroding sponge abundances on reefs, it is important to understand the factors that influence their distributions and assemblage composition. As bioeroding sponges are dependent on calcareous substrates, the availability of suitable substrate has been identified as a key factor influencing their abundance (Chaves-Fonnegra *et al.*, 2007; Carballo *et al.*, 2008; Schönberg & Ortiz, 2009; Schönberg, 2015a). Other important environmental and abiotic factors that have been previously shown to influence bioeroding sponge abundance include eutrophication (e.g. Holmes *et al.*, 2000; Chaves-Fonnegra *et al.*, 2007; Nava *et al.*, 2014), light availability (e.g. López-Victoria & Zea, 2005), and sedimentation and turbidity (e.g. Muricy, 1991; Nava & Carballo, 2013; see summary in Schönberg, 2008). Many of these environmental parameters can also be associated with reef degradation. Previous studies have shown that eutrophication and high chlorophyll concentrations are associated with low coral recruitment and species richness, and promote macroalgal abundance (e.g. De'ath & Fabricius, 2010; Fabricius, 2011). In addition to eutrophication, reefs that are subject to excessive terrestrial run-off are typically characterized by highly turbid water with high sedimentation rates. High turbidity reduces light availability for photosymbiotic hard corals and has been associated with reduced coral growth rates (Crabbe & Smith, 2005), reduced coral diversity (e.g. De'ath & Fabricius, 2010) and higher disease prevalence (Pollock *et al.*, 2014). Sedimentation can further stress and kill corals through either smothering or complete burial and has been associated with large declines in coral cover and diversity at impacted sites (see Fabricius, 2005; Erftemeijer *et al.*, 2012 for reviews).

In this study we hypothesized that coral mortality, and subsequent increases in the availability of dead calcareous substrate, promotes bioeroding sponge abundance. Furthermore, we hypothesized that bioeroding sponges are resilient to the environmental stressors (specifically eutrophication, turbidity and sedimentation) that can contribute towards coral mortality. To test this hypothesis, we investigated the influence of different biotic and abiotic factors on the abundance and assemblage composition of bioeroding sponges on Indonesian reefs. Recognizing related patterns will provide an increased understanding of how reef degradation may affect bioeroding sponge abundances and how future reefs might function if degradation continues.

## MATERIALS AND METHODS

### Study area

This study was conducted within the Wakatobi Marine National Park (Wakatobi) in south-east Sulawesi, Indonesia.

The Wakatobi supports a high diversity of marine species and was gazetted as a Biosphere Reserve by UNESCO in 2002. The region is also home to over 90,000 people who depend on the local reefs for food and other resources (Cullen *et al.*, 2007).

Surveys were conducted at seven sites on the fringing reefs around the islands of Hoga and Kaledupa in March–April 2014, and then continued in June 2014 to include another four sites around the islands of Wanci and Tomia (Figure 1). Survey sites were chosen to represent a variety of reef types, environmental conditions and levels of reef degradation. The initial seven 'core' sites included three steep-wall reefs (>70° inclination): Ridge 1 and 3, and Ridge 2; three sloping reefs (50–70°): Kaledupa 1, Kaledupa Double Spur and Pak Kasims; and Sampela 1, a more gently sloping reef adjacent to a stilted Bajo village, which is considered to be highly degraded, sedimented and turbid (Bell & Smith, 2004; McMellor & Smith, 2010; Powell *et al.*, 2014). The additional four sites were again gently sloping reefs: Karang Gurita, a coral atoll located off the island of Wanci; Tomia 1 and 2 off the island coast of Tomia, which appear relatively healthy with high cover of branching corals; and Wanci Harbour, a turbid and sedimented reef within Wanci Harbour.

### Sponge species identification

Initial surveys were conducted across the seven core sites between depths of 3–20 m, to identify the locally dominant bioeroding sponge species. Bioeroding sponges are difficult to identify *in situ*, and new studies in previously un-surveyed regions have often discovered new species (e.g. Calcinaï *et al.*, 2005). Sponges encountered during these surveys were initially identified as different operational taxonomic units (OTUs) based on recognizable external morphology and erosion chamber macro-characteristics. To create a functional context, pulse amplitude modulated fluorometry (DIVING-PAM with red excitation light, Walz, Ifeltrich) was used to detect photoactivity of *in situ* light-adapted sponges by measuring fluorescence yield; subsequent tissue examination then identified the presence of symbiotic *Symbiodinium*. Eight different OTUs were designated in this manner and three ethanol-preserved representative samples of each were transported back to the Victoria University of Wellington for species confirmation using spicule analysis. Full taxonomic results will be published elsewhere.

### Bioeroding sponge abundance and benthic composition

Bioeroding sponge abundances and substrate characteristics were assessed *in situ* using line intercept transects. Surveys were conducted at 5 and 10 m depths for each of the initial seven 'core' sites in March–April 2014. Due to logistical constraints, surveys were conducted at just 10 m depth for the four additional sites surveyed in June 2014. At each site and depth, six 10 m transects were haphazardly deployed running parallel to the reef contours with a minimum separation of 10 m between replicate surveys. Sponge abundances and substrate composition were recorded by visually inspecting ten 1 m subsamples on each transect; along each transect subsample, the total linear distance of each sponge species and

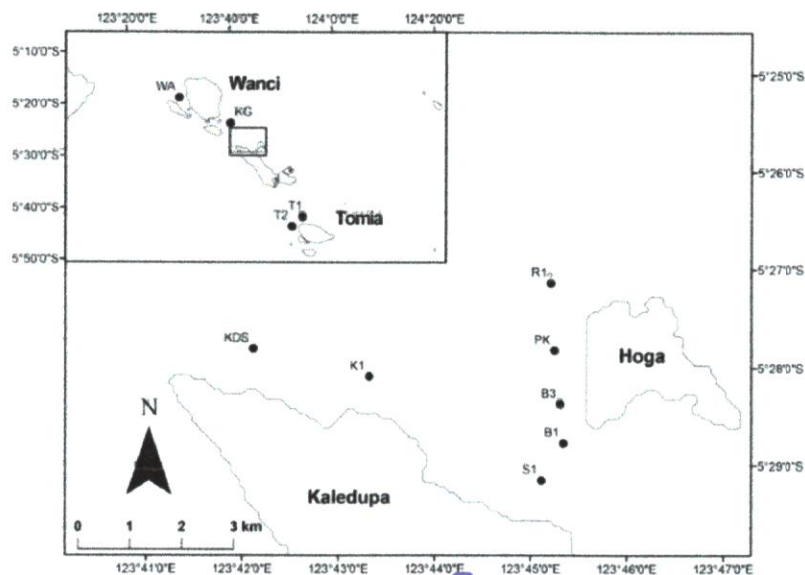


Fig. 1. Site map of the survey region in the Wakatobi (top left) and the core survey sites around the islands of Hoga and Kaledupa. Sites Buoy 1 and 3, Kaledupa 1 and Kaledupa Double Spur, Karang Gurita, Pak Kasim, Ridge 1, Sampela 1, Tomia 1 and 2, and Wanci Harbour are abbreviated as B1, B3, K1, KDS, KG, PK, R1, S1, T1, T2 and WA, respectively.

substrate type that intercepted the measuring tape were recorded (after English *et al.*, 1994; Schönberg, 2015a). Schönberg (2015a) found linear distance to be proportional to areal extent, therefore our linear distributions were treated as a proxy for percentage benthic cover. Substrate types were listed as 'live coral', 'live other', 'dead substrate' and 'sand'. The dead exposed substrate category included recently dead coral, coral rubble or calcareous platform that was uncolonized by other taxa, except for short filamentous algal turf.

### Environmental variables

Environmental conditions at each site were characterized in terms of chl-*a* concentration, turbidity, sedimentation and water flow (for full details of methods see Marlow *et al.*, in press). Average chl-*a* concentration and turbidity levels for the seven core sites were determined from multiple deployments of a XR-420 multi-channel data logger in 2010, 2014 and 2015. Due to time constraints, no chl-*a* or turbidity data were collected from the four additional sites around Tomia and Wanci. Sediment deposition and retention were indirectly evaluated in March and June 2014 by measuring depth of 'settled sediment' along 30 m transects at each site and depth (following Bell & Turner, 2000). Mean proportional current velocity for the seven core sites was quantified in June–August 2014 using multiple 24 h deployments of plaster of Paris casts, commonly referred to as 'clods' (Doty, 1971; Jokiel & Morrissey, 1993).

### Statistical analysis

All univariate statistical analyses were performed in SPSS (version 21; IBM) and multivariate analysis using the

PRIMER-E v6 software package with the permutational multivariate analysis of variance (PERMANOVA) add on (Clarke & Gorley, 2006).

The average abundance of each bioeroding sponge species across each transect was calculated from the linear extent of each of the ten 1 m subsamples and used as a proportional indicator for percentage cover (see Schönberg, 2015a). Previous studies have consistently found that bioeroding sponge abundance correlates with the availability of dead calcareous substrate (Chaves-Fonnegra *et al.*, 2007; Carballo *et al.*, 2008; Schönberg & Ortiz, 2009). Therefore analysis of environmental drivers of bioeroding sponge abundance should use abundance data that are standardized to the local availability of dead substrate (Schönberg, 2015a). After finding similar correlations in the Wakatobi (see below), total bioeroding sponge abundance and individual species abundances were standardized to available dead substrate (including that which the sponges already occupy).

The lack of chl-*a*, turbidity or water movement data from the four additional sites meant that the analysis of interactions between environmental variables and boring sponges only incorporated data from the core sites. Due to the considerable differences in environmental conditions between Sampela 1 and the other sites, and the tendency for this site to drive collinearity amongst variables, the analysis of the core sites was conducted twice, once including Sampela 1 and once without.

### BIOERODING SPONGE ABUNDANCE AND DIVERSITY

Differences in total bioeroding sponge abundance and species diversity (Shannon's index;  $H'$ ) between sites and depths were tested using a two factorial general linear model (GLM).

Where significant differences were found Tukey's or Gabriel's (in the case of unequal sample sizes) *post hoc* procedures were used to identify where the differences occurred. The influence of individual environmental/benthic factors on bioeroding sponge abundance and diversity were assessed with Pearson's product-moment correlation coefficients. To determine whether these same factors constituted stressors for corals, environmental/benthic data was also correlated against coral cover using Pearson's product-moment correlation coefficients. Data were square-root or log transformed to meet the necessary assumptions of equal variance and normality for both tests. For any data that failed to meet these assumptions despite transformation, non-parametric tests were used (a Kruskal–Wallis or Spearman's Rank test).

## ENVIRONMENTAL CHARACTERIZATION

Environmental characterization of each site was achieved using a multivariate analysis within PRIMER. All analyses were based on resemblance matrices calculated from normalized data using Euclidean-Distance similarity coefficients. Draftman's plots of all pairwise correlations were checked for collinearity and skewness in the abiotic and biotic data; these suggested that fourth-root transformation was appropriate for an increase in normality for all the variables. Substrate and environmental differences between sites and depths were examined using one and two-factor PERMANOVAs (site and depth as factors with 7 levels or site as factor with 11 levels), and results were graphically displayed through principal coordinate analysis (PCO). The inter-site differences in abiotic and biotic variables were characterized with Pearson's correlations ( $>0.4$ ) between PCO axes and the individual abiotic/biotic components.

## BIOERODING SPONGE ASSEMBLAGE PATTERNS

All biological assemblage data analyses were based on resemblance matrices using Bray–Curtis similarity coefficients calculated from square-root transformed data. Due to the general sparseness or patchiness of bioeroding sponge abundances and the complete absence on one transect, a dummy species with the abundance value of 0.5 was added to each transect in order to diminish the erratic behaviour of the Bray–Curtis coefficient (Clarke *et al.*, 2006). To identify those species that were contributing towards any assemblage dissimilarity between sites or depths, a SIMPER analysis was performed. Associations between bioeroding sponge assemblage structure and abiotic/biotic variables were investigated in a distance-based multiple linear regression model (DISTLM), which is a non-parametric permutation-based procedure that enables significance testing of explanatory variables against multivariate response variables (Anderson *et al.*, 2008). In order to find the most parsimonious model, Akaike's Information Criterion (AIC), a step-wise procedure, adjusted for small sample sizes (AICc), was used (Burnham & Anderson, 2004). As with univariate correlations, the DISTLMs were repeated twice, once with the outlier site Sampela 1 and once without. DISTLM outputs were represented graphically with a distance-based redundancy

ordination (dbRDA), including a 2D scatterplot to identify differences between sites and 2D bubble plots to identify differences in species abundances. The influence of the predictor variables selected by the DISTLM on the abundance of individual species that were contributing the greatest difference between sites were tested with Spearman's Rank correlations.

## RESULTS

### Bioeroding sponges

#### TAXA PRESENT

The Wakatobi bioeroding sponge fauna consisted of eight distinct species. Six of these sponges were characterized at the species level: *Cliona cf. schmidtii* (Ridley, 1881), *Cliona orientalis* Thiele, 1900, *Cliothosa hancocki* (Topsent, 1888), *Cliothosa cf. aurivillii* (Lindgren, 1897), *Spheciospongia cf. vagabunda* (Ridley, 1884) and *Zyzzocriceta* Schönberg, 2000. Two species were allocated to the *Cliona viridis* (Schmidt, 1862) species complex, but were not identified further (*Cliona aff. viridis* sp. A & B). The abbreviations cf. and aff. are used to indicate that a species is either similar and possibly conspecific (cf.) or similar but likely different (aff.) to a previously described species. Three species, *C. orientalis* and *C. aff. viridis* sp. A and B, were hosts to photosynthetic dinoflagellates (*Symbiodinium* spp.).

#### ABUNDANCE, DISTRIBUTION AND DIVERSITY

Bioeroding sponges occupied an average of 3.1% ( $\pm 0.21$  SE) of the total reef area, and total abundance varied significantly among sites (GLM,  $F_{(10,108)} = 5.721$ ,  $P < 0.001$ ), but not between the depths of 5 and 10 m (GLM,  $F_{(1,108)} = 0.242$ ,  $P = 0.624$ ). Abundance was highest at Wanci Harbour (4.9% of total reef area  $\pm 0.63$  SE) and lowest at Tomia 1 (1.3% of total reef area  $\pm 0.31$  SE).

After standardizing sponge abundance to dead substrate availability (standardized bioeroding sponge abundance herein referred to as % DS), bioeroding sponges were found to occupy an average of 8.9% DS ( $\pm 0.7$  SE) across all sites. There was a significant difference between sites (GLM,  $F_{(10,108)} = 17.366$ ,  $P < 0.001$ ) with up to 21.9% DS ( $\pm 2.7$  SE) at Buoy 3 and as low as 3.5% DS ( $\pm 0.8$  SE) at Sampela 1 (Figure 2A). There was also a significant difference between depths (GLM,  $F_{(10,108)} = 10.445$ ,  $P = 0.002$ ) with 9.7% DS ( $\pm 0.5$  SE) at 10 m depth and 7.7% DS ( $\pm 0.6$  SE) at 5 m depth.

Species diversity of the bioeroding sponges differed significantly among the sites, but not between depths (Kruskal–Wallis,  $P = 0.003$  and  $P = 0.986$ , respectively). Species diversity was highest at Wanci Harbour and Buoys 1 & 3 ( $H' = 1.173$ , 1.098 and 1.102, respectively), and lowest at Karang Gurita and Tomia 1 ( $H' = 0.380$  and 0.385, respectively; Figure 2B).

#### ASSEMBLAGE COMPOSITION

Although the field surveys recognized eight bioeroding sponge species within the Wakatobi assemblage, 81.8% of the abundance of the sponges was represented by only three species: *C. aff. viridis* sp. B (39.1%), *Cl. cf. aurivillii* (23.2%) and *S. cf. vagabunda* (19.5%). Sponge assemblage composition was significantly different among sites (PERMANOVA,

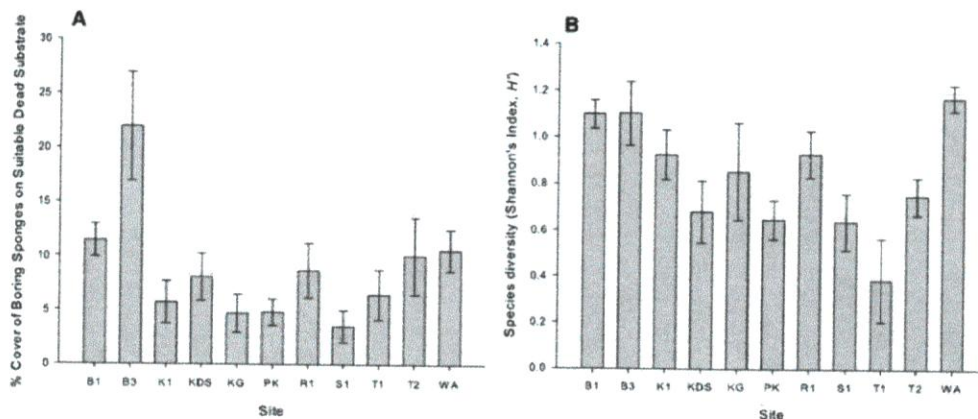


Fig. 2. Bioeroding sponge distribution patterns per sample site and across all depths. (A) Total sponge abundance (standardized to substrate). (B) Species diversity. Bar height represents mean  $\pm$  SE. See Figure 1 for site abbreviations.

Pseudo- $F = 5.1843$ ,  $P = 0.001$ ; Figure 3), and between 5 and 10 m depths (PERMANOVA, Pseudo- $F = 6.3$ ,  $P = 0.001$ ). Given this influence of depth on assemblage composition, further multivariate analysis was restricted to the seven core sites, which included survey data from both depth categories.

The three largest inter-site differences in assemblage composition occurred between Buoy 3 vs Sampela 1, Kaledupa Double Spur and Kaledupa 1 (80.0, 70.1 and 68.6%, respectively), and were largely driven by differences in the abundance of *S. cf. vagabunda*. This species was relatively abundant at Buoy 3 (average abundance of 13.3% DS and within-site assemblage similarity contribution of 58.0%), but had a low abundance at Sampela 1 (<0.1% DS), Kaledupa Double Spur (0.7% DS) and Kaledupa 1 (0.5% DS), contributing 40.7, 35.3 and 37.8% towards inter-site assemblage dissimilarity, respectively.

The majority (68.1%) of the 52.6% difference in species composition between depths was attributed to depth-related variation in the abundance of three most common species, *C. aff. viridis* sp. B, *Cl. cf. aurivillii* and *S. cf. vagabunda*. Variation in *Cl. cf. aurivillii* abundance contributed 25.7% of the assemblage difference; average abundance (2.6% DS) was higher at 5 m than at 10 m (1.1% DS). The reverse was true for *S. cf. vagabunda*; average abundance at 5 m (1.5% DS) was almost threefold lower than at 10 m (4.3% DS), accounting for 19.1% of the variation in assemblage composition between depths. Variation in the abundance of *C. aff. viridis* sp. B accounted for 23.4% of variation, and although average abundance was similar at 5 and 10 m depths (2.6 and 3.1% DS, respectively), there was high within-group variation.

## Environment

### SUBSTRATE

Substrate composition did not differ significantly with depth (PERMANOVA, Pseudo- $F = 1.7118$ ,  $P = 0.143$ ) but differed significantly among the sites (PERMANOVA, Pseudo- $F = 9.1056$ ,  $P = 0.001$ ; Table 1), with 59.2% of the variation being explained by the principal coordinate analysis (PCO);

Figure 4A). Tomia 1 and Buoy 3 were characterized by hard corals; Kaledupa Double Spur and Kaledupa 1 by dead substrate; Karang Gurita, Sampela 1 and Wanci Harbour by sand; and the other four sites were more heterogeneous in their substrate composition (Figure 4A; Table 1).

### CHL-A, TURBIDITY, SETTLED SEDIMENT AND WATER FLOW

Environmental conditions (chl-*a*, turbidity, settled sediment, water flow) varied significantly among the core sites (PERMANOVA, Pseudo- $F = 26.572$ ,  $P = 0.001$ ), but not between depths. The two axes of the PCO together explained 63.4% of the variation, further characterizing the core sites by demonstrating: elevated chl-*a* concentration, settled sediment and turbidity at Sampela 1; low flow rates at Buoys 1 and 3; high flow rates at Kaledupa 1, Kaledupa Double Spur and Ridge 1; and comparatively heterogeneous environmental conditions at Pak Kasim (Figure 4B; Table 1).

## How did the environment affect bioeroding sponges?

### SPONGE ABUNDANCE AND DIVERSITY

Overall, total bioeroding sponge abundance was positively correlated with availability of dead calcareous substrate (Pearson's,  $r = 0.345$ ,  $P < 0.001$ ), negatively correlated with the abundance of sandy substrate (Pearson's,  $r = -0.199$ ,  $P < 0.039$ ), and showed no significant correlation with either live coral or other live cover (Table 2).

Abundance (% DS) decreased on reefs with higher water movement and on those with high chl-*a* concentrations (Table 2). Omitting Sampela 1, data still showed that within the remaining core sites water movement correlated negatively with % DS, but greater depth of settled sediment and turbidity were positively correlated with % DS and chl-*a* concentration had no influence (Table 2). The abundance of different substrate types (with the exception of dead substrate) also showed no significant correlation with % DS of these sponges (Table 2).



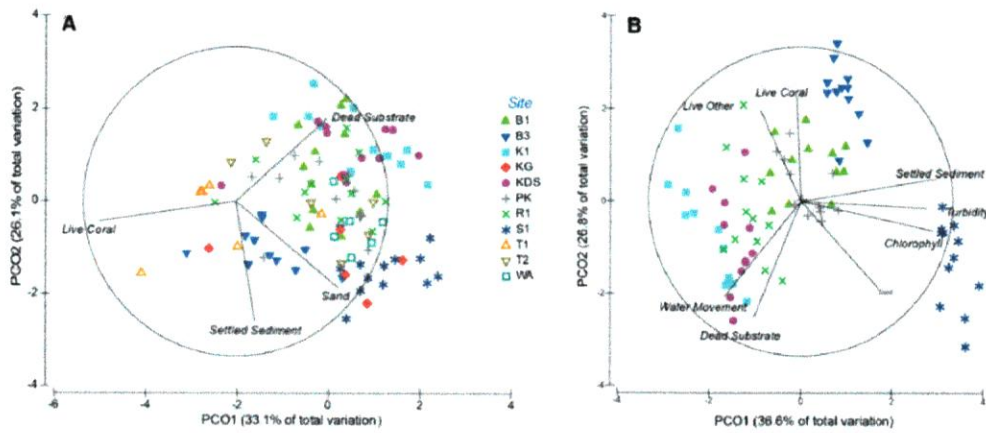


Fig. 4. Two dimensional representation of site similarities with respect to their multivariate physical and water quality parameters (principal coordinate analysis, PCO). (A) Comparison of all 11 study sites with regards to substrate composition. (B) Comparison between the seven core study sites including substrate composition and selected environmental factors as chosen for this study. Overlaid vectors represent components that have a Pearson's correlation of greater than 0.4 with either of the PCO axes. See Figure 1 for site abbreviations.

Sponge species diversity increased with increasing proportional contribution of 'other live' benthos, regardless of whether or not Sampela 1 was included in the analysis. However, a significant negative correlation with chl-*a* concentration only occurred when Sampela 1 was included in the analysis, and a significant negative correlation with turbidity only when Sampela 1 was omitted (Table 2).

#### ASSEMBLAGE COMPOSITION

The best sponge assemblage model in DISTLM explained 30.1% of variation in assemblage structure based on non-substrate-standardized abundance: dead substrate (14.0%), chl-*a* concentration (6.4%), water movement (4.9%) and our proxy for sedimentation, thickness of settled sediment (4.9%) (Figure 5A). Removal of the Sampela 1 data increased the total explanation value slightly to 33.0%. The explanatory power of dead substrate (16.1%), water movement (5.2%) and settled sediment (4.0%) changed little, there was no explanatory value for chl-*a* concentration, and changes in turbidity (4.2%) and live coral cover (3.5%) were found to have only slight influence (Figure 5B).

When abundance was standardized to dead substrate availability, 23.9% of assemblage structure variation was explained by environmental variables including water movement (9.3%), settled sediment (8.2%) and chl-*a* concentration (6.4%). The removal of Sampela 1 from this analysis explained a similar amount of variation (24.4%), but the explanatory value of settled sediment was greater (14.8%), water movement became less important (4.4%), chl-*a* concentration no longer had an effect, and turbidity was found to be more important (5.2%).

#### INDIVIDUAL SPECIES

The abundance of dead substrate correlated positively with two of the three dominant boring sponge species: *C. cf. viridis* sp. B and *Cl. cf. aurivillii* (Spearman's Rank,  $r_s = 0.245$ ,  $P = 0.011$  and  $r_s = 0.552$ ,  $P < 0.001$ , respectively; Figure 6). However, the abundance of *S. cf. vagabunda* had

an inverse relationship with dead substrate (Spearman's Rank,  $r_s = -0.299$ ,  $P = 0.002$ ) (Figure 6). When the abundance of these species was standardized to dead substrate availability their individual relationships with the explanatory variables identified by the DISTLM were mixed. None of the variables correlated with *C. aff. viridis* sp. B % DS, *Cl. cf. aurivillii* % DS correlated negatively with the depth of settled sediment, and *S. cf. vagabunda* % DS correlated positively with the depth of settled sediment but negatively with water movement (Appendix). Omitting Sampela 1 changed these relationships very little, but revealed a stronger relationship between *S. cf. vagabunda* % DS and settled sediment, as well as a positive correlation with turbidity (Appendix).

#### HARD CORALS

Hard coral cover was highest at Tomia 1 (64.58% of total reef area  $\pm 7.05$ ) and lowest at Kaledupa 1 and Sampela 1 (11%  $\pm 1.79$  and 39.58%  $\pm 2.27$  of total reef area, respectively; Table 2). Coral cover was significantly negatively correlated with the abundance of dead substrate, live 'other', sand and water movement (Table 2). When omitting Sampela 1 from the analysis, coral cover remained negatively correlated with all these factors, but a positive correlation with chl-*a* also emerged (Table 2).

#### DISCUSSION

By identifying putative environmental drivers of bioeroding sponge abundance, this study provided an opportunity to assess how different bioeroding sponges respond to environmental variability using a model reef system. We hypothesized that factors that can lead to coral mortality result in increased availability of dead calcareous substrate, and that such factors are thus beneficial for bioeroding sponges. Our results showed that dead substrate availability was greater on reefs with low coral cover. Furthermore, the availability of this suitable dead substrate was correlated with overall bioeroding

Table 1. Summary table of substrate and environmental characteristics of each site. Standard errors shown. See Figure 1 for site abbreviations. Percentages refer to proportional linear extent along intercept transects.

| Factor             | Unit                | Site | B1            | B3            | K1            | KDS           | KG            | PK            | R1            | S1            | T1            | T2             | WA            |
|--------------------|---------------------|------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|----------------|---------------|
| Live coral         | %                   |      | 19.29 (±2.57) | 35.88 (±4.20) | 11 (±1.79)    | 15.04 (±4.06) | 19.25 (±8.91) | 23.75 (±4.30) | 27.98 (±4.55) | 11.58 (±2.27) | 64.58 (±7.05) | 27.58 (±8.41)  | 18.0 (±2.55)  |
| Dead substrate     | %                   |      | 37.96 (±3.80) | 16.17 (±1.80) | 39.98 (±4.07) | 46.04 (±4.94) | 24.33 (±4.53) | 33.63 (±2.16) | 41.17 (±4.05) | 34.08 (±3.04) | 20.33 (±4.96) | 28.42 (±4.58)  | 41.92 (±4.41) |
| Live other         | %                   |      | 18.04 (±3.22) | 45.12 (±3.35) | 35.13 (±4.82) | 35.38 (±4.50) | 36.33 (±7.64) | 33.91 (±3.76) | 24.92 (±2.61) | 23.25 (±1.84) | 10.75 (±1.17) | 14.42 (±1.93)  | 31.92 (±3.57) |
| Sand               | %                   |      | 4.52 (±1.46)  | 4.21 (±1.92)  | 7.67 (±2.18)  | 3.88 (±1.48)  | 20.08 (±9.34) | 8.29 (±2.94)  | 8.39 (±3.39)  | 30.38 (±3.54) | 4.42 (±1.05)  | 29.42 (±11.84) | 7.67 (±2.66)  |
| Sediment thickness | mm                  |      | 0.78 (±0.18)  | 1.95 (±0.37)  | 0.25 (±0.12)  | 0.5 (±0.10)   | 2.23 (±0.54)  | 1.06 (±0.13)  | 0.85 (±0.16)  | 2.94 (±0.28)  | 0.5 (±0.17)   | 0.4 (±0.12)    | 2.45 (±0.86)  |
| Water movement     | % cloud weight loss |      | 8.36 (±0.84)  | 8.79 (±0.75)  | 18.73 (±0.67) | 24.42 (±1.81) | n/a           | 12.78 (±2.60) | 16.93 (±1.62) | 13.88 (±0.81) | n/a           | n/a            | n/a           |
| Turbidity          | NTU                 |      | 2.66 (±1.22)  | 2.06 (±0.7)   | 1.69 (±0.5)   | 2.07 (±0.4)   | n/a           | 1.75 (±0.2)   | 1.32 (±0.6)   | 4.10 (±1.04)  | n/a           | n/a            | n/a           |
| Chl- <i>a</i>      | µg l <sup>-1</sup>  |      | 0.36 (±0.12)  | 0.36 (±0.02)  | 0.12 (±0.1)   | 0.31 (±0.08)  | n/a           | 0.94 (±0.14)  | 0.33 (±0.03)  | 2.54 (±1.00)  | n/a           | n/a            | n/a           |

Table 2. Results of bivariate correlations comparing total sponge abundance (TA), substrate standardized abundance (STA), bioeroding sponge species diversity (SD) and coral cover (CC) with substrate characteristics and environmental factors. Abundance correlations using Pearson's and species diversity correlations using Spearman's Rank.

| Factor           | Dependent variable | Sampela included |                  | Sampela excluded |                  |
|------------------|--------------------|------------------|------------------|------------------|------------------|
|                  |                    | r/r <sub>s</sub> | P                | r/r <sub>s</sub> | P                |
| Live coral       | TA                 | -0.159           | 0.099            | <b>-0.287</b>    | <b>0.005</b>     |
|                  | STA                | 0.186            | 0.054            | 0.031            | 0.765            |
|                  | SD                 | 0.048            | 0.622            | -0.033           | 0.750            |
|                  | CC                 | -                | -                | -                | -                |
| Live other       | TA                 | 0.055            | 0.571            | 0.080            | 0.436            |
|                  | STA                | 0.164            | 0.090            | <b>0.219</b>     | <b>0.032</b>     |
|                  | SD                 | <b>0.206</b>     | <b>0.032</b>     | <b>0.336</b>     | <b>0.021</b>     |
|                  | CC                 | -0.243           | 0.011            | -0.315           | 0.002            |
| Dead substrate   | TA                 | <b>0.345</b>     | <b>&lt;0.001</b> | <b>0.344</b>     | <b>0.001</b>     |
|                  | STA                | -                | -                | -                | -                |
|                  | SD                 | 0.152            | 0.117            | 0.134            | 0.194            |
|                  | CC                 | <b>-0.406</b>    | <b>&lt;0.001</b> | <b>-0.445</b>    | <b>&lt;0.001</b> |
| Sand             | TA                 | <b>-0.199</b>    | <b>0.039</b>     | -0.077           | 0.455            |
|                  | STA                | -0.126           | 0.1944           | 0.062            | 0.547            |
|                  | SD                 | -0.073           | 0.451            | 0.019            | 0.855            |
|                  | CC                 | -0.449           | <b>&lt;0.001</b> | -0.390           | <b>&lt;0.001</b> |
| Settled sediment | TA                 | -0.110           | 0.256            | 0.176            | 0.087            |
|                  | STA                | -0.950           | 0.388            | <b>0.483</b>     | <b>&lt;0.001</b> |
|                  | SD                 | -0.036           | 0.712            | 0.081            | 0.432            |
|                  | CC                 | -0.054           | 0.577            | 0.149            | 0.147            |
| Turbidity        | TA                 | <b>-0.364</b>    | <b>0.001</b>     | <b>0.306</b>     | <b>0.009</b>     |
|                  | STA                | -0.177           | 0.107            | <b>0.471</b>     | <b>&lt;0.001</b> |
|                  | SD                 | 0.070            | -0.527           | <b>0.309</b>     | <b>0.008</b>     |
|                  | CC                 | -0.211           | 0.054            | 0.080            | 0.503            |
| Chl- <i>a</i>    | TA                 | <b>-0.400</b>    | <b>&lt;0.001</b> | -0.222           | 0.061            |
|                  | STA                | <b>-0.440</b>    | <b>&lt;0.001</b> | -0.159           | 0.183            |
|                  | SD                 | <b>-0.277</b>    | <b>0.011</b>     | -0.161           | 0.177            |
|                  | CC                 | -0.112           | 0.309            | <b>0.316</b>     | <b>0.007</b>     |
| Water movement   | TA                 | -0.191           | 0.082            | -0.189           | 0.111            |
|                  | STA                | <b>-0.328</b>    | <b>0.002</b>     | <b>-0.435</b>    | <b>&lt;0.001</b> |
|                  | SD                 | -0.061           | 0.583            | -0.097           | 0.415            |
|                  | CC                 | -0.323           | 0.003            | -0.392           | 0.001            |

Significant correlations in bold.

sponge abundance, and was the best predictor of differences in assemblage composition. However, bioeroding sponge abundance was lowest on a sedimented reef, despite high availability of dead substrate, demonstrating a limited resilience to sedimentation in some bioeroding sponge species. This indicates that not all forms of coral mortality, and subsequent increases in dead substrate availability, are beneficial for bioeroding sponges. Finally, inter-species variation in abundance with respect to environmental factors illustrated how largely ubiquitous patterns cannot easily be generalized across the entire taxon group.

### Abiotic and biotic influences on bioeroding sponges

The availability of dead calcareous substrate was one of the most important factors causing differences in bioeroding sponge abundance and assemblage composition on Wakatobi reefs. This is consistent with earlier studies in Indonesia (Holmes *et al.*, 2000), on the Australian GBR (Schönberg,

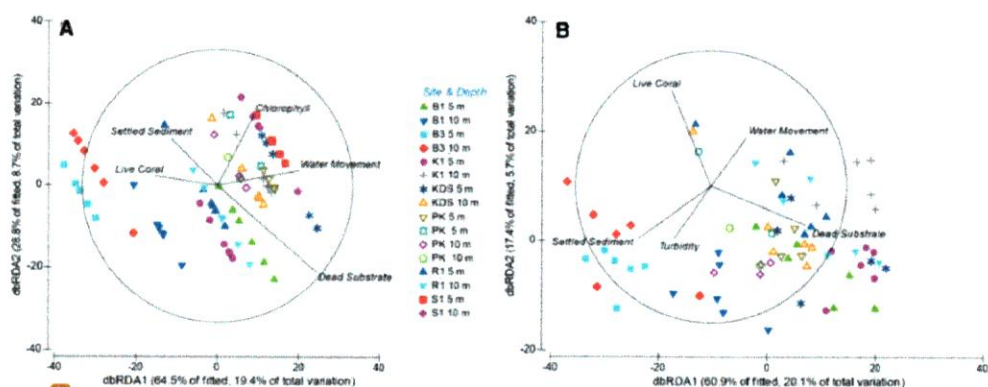


Fig. 5. Distance based redundancy analysis (dbRDA) ordinations of Bray-Curtis models for assemblage composition. (A) Seven core sites and both depth categories. (B) Core sites and both depth categories with the omission of Sampela 1. See Figure 3 for site abbreviations.

2001, 2015a; Schönberg & Ortiz, 2009), in the Mexican Pacific (Va & Carballo, 2013) and in the Caribbean (Callahan, 2005; Chaves-Fonnegra *et al.*, 2007). This observation is a consequence of the sponges' endolithic habit that creates a dependency on calcareous structures (e.g. Schönberg, 2008). Nevertheless, this relationship was not uniform across all species studied, and the association was primarily observed in the papillate *Clathrospira cf. aurivillii* and *Cliona aff. viridis* sp. B. Both species dominated at three sites with a high occurrence of dead substrate and little live coral. Conversely, *Speciospongia cf. vagabunda* was more common where dead substrate availability was low, but coral cover was relatively high. This suggests that biological requirements vary between these species, which could be due to different levels of dependency on endolithic life style. Some *Speciospongia* spp. are facultative endoliths, known to develop into fleshy or massive morphologies that are largely independent of the endolithic habit (Schönberg *et al.*, 2017b). *Cl. aurivillii* was originally described as occurring in a free-living, as well as in an endolithic form (Lindgren, 1897), but recent reports only observed the endolithic form (e.g. Calcina *et al.*, 2006). We do not know enough about *Cliona aff. viridis* sp. B to assess how tightly it is associated with calcareous substrates, but in Wakatobi it was only observed in endolithic form. Furthermore, dispersal capabilities of bioeroding sponges are thought to vary significantly, from the extremely limited (<10 m) to the extensive (>10 km) (Mariani *et al.*, 2000; Bautista-Guerrero *et al.*, 2010, 2016; Chaves-Fonnegra *et al.*, 2015). Thus, depending on species-specific traits, distribution patterns can be further decoupled from substrate availability, as some recruits may settle close to the parent sponge rather than evenly spreading out across all available substrate (Callahan, 2005).

As many bioeroding sponges are heterotrophic (or at least partly), they are reliant on particulate and dissolved organic carbon to meet their energetic demands (Mueller *et al.*, 2014). Nutrient availability, typically measured as chl-*a*, has consequently repeatedly been shown to correlate with bioeroding sponge abundance and erosion rates (e.g. Rose & Risk, 1985; Holmes *et al.*, 2000; Holmes *et al.*, 2009). However, in our study we found no correlation between bioeroding sponge abundance and chl-*a* concentration when data

from Sampela 1 was excluded from the analysis. The lack of a clear nutrient stimulus may be explained by the generally low and similar chl-*a* concentrations at most sites (other than Sampela 1). Similar results have been found elsewhere in Indonesia; Holmes *et al.* (2000) found that bioeroding sponge occupation of massive corals correlated with chl-*a* concentration on polluted Javan reefs but not in Ambon where chl-*a* was less concentrated and more consistent across sites. The majority of our chl-*a* measurements deployments took place during the dry season (June–August) when chl-*a* concentration is at its highest (Tadjudah *et al.*, 2012). However, the broad scale seasonal upwellings that are responsible for this peak in chl-*a* (Wyrski, 1961), might also mask inter-site variation in chl-*a* concentration from local sources. These are likely to be more pronounced in the wet season when chl-*a* levels are often higher on coastal reefs (e.g. Lapointe *et al.*, 2004; Devlin *et al.*, 2013). The negative correlation between total bioeroding sponge abundance and chl-*a* concentration, when including Sampela 1 (which had the highest chl-*a* concentration), appears inconsistent with the concept of nutrient enrichment being beneficial to these sponges. However, Sampela 1 was not only characterized by high chl-*a* levels, we found it to be the most turbid and sedimented of the surveyed sites (as is consistent with previous descriptions of this reef (see Crabbe & Smith, 2002, 2005; Hennige *et al.*, 2008, 2010; Biggerstaff *et al.*, 2015, 2017)). In these circumstances, interactions with other factors, such as sedimentation, can mask the effects of nutrient enrichment, leading to different net responses (e.g. Holmes *et al.*, 2000; Reis & Leão, 2000; Chaves-Fonnegra *et al.*, 2007; Holmes *et al.*, 2009; Nava *et al.*, 2014). Low abundances of bioeroding sponges in eutrophic environments that experience excessive sedimentation or suspended particle concentrations have also been found in other studies. For example, Edinger *et al.* (2000) noted that bioeroding sponges were rare on reefs in the highly eutrophic Jakarta Bay, which the authors attributed to high sedimentation levels. Nava & Carballo (2013) came to a similar conclusion when they observed no relationship between the distribution of boring sponges and chl-*a* concentration on Pacific Mexican reefs.

Sedimentation can influence sponge distribution patterns by preventing settlement, by impeding vital physiological

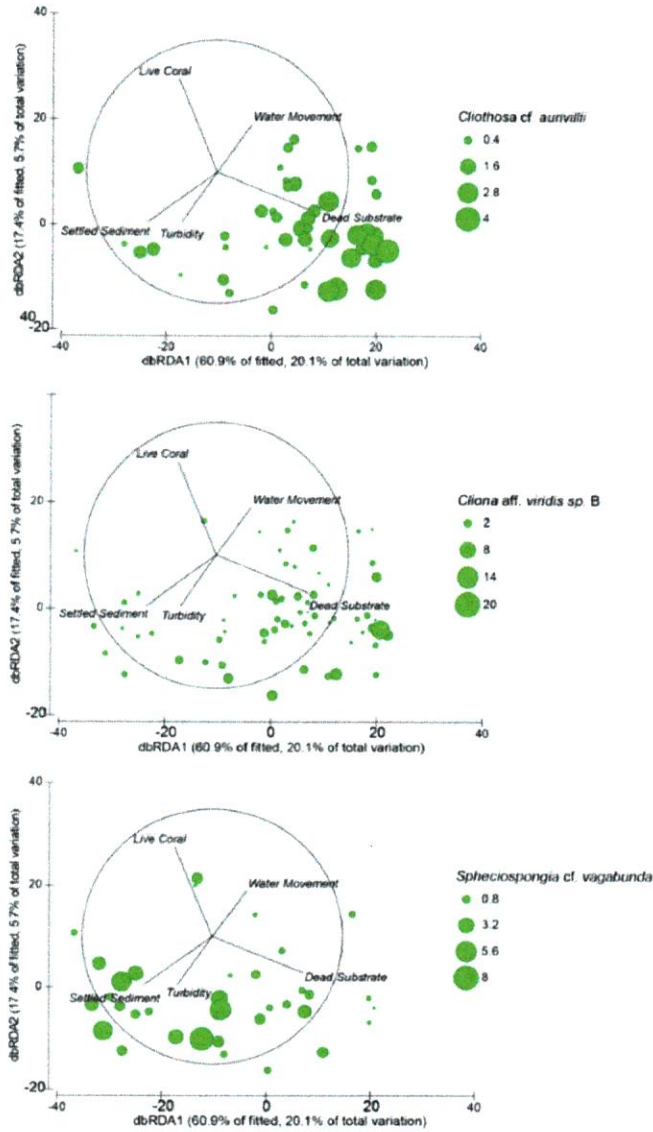


Fig. 6. Distance based redundancy analysis (dbRDA) ordinations of fitted models for assemblage composition of seven core sites (both depth categories), with the omission of Sampela 1. Overlapping 2D bubbles represent the abundance *Clotrocha cf. aurivillii* (top), *Cliona aff. viridis sp. B* (middle) and *Spherospongia cf. vagabunda* (bottom).

functions or by controlling survival and therefore selecting for adapted species over others (Bell *et al.*, 2015; Schönberg, 2016a, b). Bioeroding sponge assemblage compositions in sedimented environments are consequently structured by the respective differences in species sediment tolerance (Carballo *et al.*, 1994; Nava & Carballo, 2013). Some bioeroding species are particularly tolerant of high levels of sediment deposition, becoming covered by a thin layer of sediment, or are even adapted to live mostly buried in sediments

(endopsammic; Schönberg, 2016a). Sediment tolerance mechanisms in bioeroding sponge include fistular projections above the sediment layer, self-cleaning surfaces and temporary oscula closure (Schönberg, 2015b, 2016a; Pineda *et al.*, 2017a). However, even these tolerant species are thought to require a patch of free substrate for larval settlement (Schönberg, 2016a). In our study the impact of sedimentation primarily emerged as reduced abundance and species richness at the sedimented site, Sampela 1. Previous work by Crabbe &

Smith (2005) found sedimentation rates were up to 3.8× higher at Sampela 1 than on other local reefs and Biggerstaff *et al.* (2017) estimated average Sampela sedimentation rates to be 4.40 (±0.30) mg cm<sup>-2</sup> d<sup>-1</sup>, which is comparable to other Indo-Pacific reefs that are considered to be highly sedimented (e.g. Golbuu *et al.*, 2011; Bannister *et al.*, 2012). One species that was absent from Sampela 1 was the endolithic, phototrophic *Cliona orientalis*. Recent observations identified this species as comparatively tolerant of sediment deposition and able to maintain clean surfaces (Büttner & Siebler, 2013; Schönberg, 2015b; Pineda *et al.*, 2017b). However, under extreme and long-term sediment pressure including high concentrations of suspended particles, *C. orientalis* experienced bleaching and mortality (Pineda *et al.*, 2017a, c), which might attest to its dependency on its dinoflagellate symbionts (Fang *et al.*, 2014, 2017). *Cliona orientalis*' sediment intolerance appears to relate to the quality of suspended particles; on the GBR it displayed more severe responses to applications of mud than sand and was less abundant in reef areas with a higher proportion of fine sediments (Büttner & Siebler, 2013; Ramsby *et al.*, 2017). Over 30% of settled sediment at Sampela 1 is comprised of particles finer than 63 µm (Biggerstaff *et al.*, 2017), suggesting that the sediment regime is locally preventing the persistence or even the establishment of *C. orientalis*. The high concentration of fine sediment could also explain the near-absence of *S. cf. vagabunda* at Sampela 1, which contradicts the positive correlation between this species' abundance and settled sediment depth. Several species within the genus *Sphaerospongia* are known to benefit from living embedded in sediments (Schönberg, 2016a) and *S. cf. vagabunda* was most abundant at Buoy 1 and 3, which had moderately high levels of sedimentation. However, previous work at Buoy 1 and 3, has shown the settled sediment composition to be much coarser than at Sampela 1 (Bell & Smith, 2004). High levels of fine sediments, suspended or deposited, seem to be a problem for many sponges, bioeroding or otherwise (Schönberg, 2001, 2016a; Schönberg & Ortiz, 2009). With little selective control over filtering intake (Reiswig, 1971), sponges are more susceptible to clogging by finer particles, thereby reducing pumping, feeding efficiency and other biological processes (see Bell *et al.*, 2015). Fine sediments are also more associated with terrestrial inputs (Bannister *et al.*, 2012) and once settled are more likely to be resuspended by currents, tides and waves (Ogston *et al.*, 2004), contributing to reef turbidity.

Low levels of particle suspension (turbidity) in the water column were beneficial for the Wakatobi bioeroding sponges and may have brought about nutrient enrichment (as per Holmes *et al.*, 2000). However, evidence from Sampela 1 suggested that excessive turbidity may have limited the abundance of bioeroding sponges as a community. Turbidity levels at the Sampela 1 are within the range that is considered stressful to corals (Cooper *et al.*, 2008) and has already been associated with low coral growth rates at Sampela 1 (Crabbe & Smith, 2005; Hennige *et al.*, 2008). Sponges also have the capacity to be negatively affected by high turbidity, either by direct impairment of the pumping and filtration apparatus or indirectly through shading (Pang, 1973; Bell *et al.*, 2015; Schönberg, 2016b). The absence of two of the three locally abundant phototrophic zooxanthellate species at the sampled depths at Sampela 1 would support the latter. On the GBR, *C. orientalis* (one of the absent species) is most abundant at ~5 m depth on clear-water reefs, but at more turbid

sample sites it dominates in the upper subtidal and intertidal regions (Bergman, 1983, as *C. viridis*; Schönberg, 2001). Extensive work on the impacts of dredging on sponges in the GBR by Pineda *et al.* (2016, 2017a, b, c) has also shown that *C. orientalis* bleaches when severely light limited and when exposed to high concentrations of suspended sediments for intermediate periods. The *Symbiodinium* within the bleached *C. orientalis* are thought to survive through parasitism of their host (Fang *et al.*, 2017) and therefore the host is unlikely to survive prolonged periods of excessive turbidity. However, light requirements are likely species specific, as other zooxanthellate clonoids appear capable of persisting in conditions of greatly reduced light availability (Rosell & Uriz, 1991).

In total no more than 33% of assemblage variability was explained by our environmental/substrate variables and no more than 16% of variability was explained by any one variable. These low explanatory values are partly a reflection of inter-species differences in environmental requirements and resilience, but are also likely due to the importance of unmeasured variables such as slope, sediment grain size or PAR (photosynthetic active radiation). Furthermore, the highest values for chl-*a*, settled sediment and turbidity all occurred at Sampela 1, likely driving collinearity and making interpretation of the impacts of these individual factors difficult. As these factors naturally occur concurrently on reefs impacted by terrestrial run-off (Fabricius, 2005), confident *in situ* analysis of the individual effects of these factors can be problematic. This is potentially best achieved using experimental manipulations. For example Pineda *et al.* (2016, 2017a, b, c) investigated the impacts of dredging on a variety of common GBR sponges (including *C. orientalis*) using separate light attenuation, settled sediment and suspended sediment based *ex situ* experiments. However as the results from our paper demonstrate, these experiments need to be tailored to the relevant species or risk extrapolation to species with different biological requirements.

#### Coral cover and stressors

Hard corals covered ~25% of reef area in the Wakatobi. This is slightly higher than the regional average (Bruno & Selig, 2007), but lower than the 32% cover recorded at similar depths in the Wakatobi in the mid-1980s (Soekarno, 1989). In the absence of appropriate time-series data it is impossible to attribute this decline to any particular source of coral degradation, but our study found no discernible impact of either sedimentation, turbidity or chl-*a* concentration on hard coral cover. This suggests that these particular coral stressors are not currently significant regulators of coral cover across the Wakatobi. The exception to this may be at a small selection of sites, such as Sampela 1. The lack of any evidence for this in our analysis may be due to the generally low levels of sedimentation, turbidity and chl-*a* across most sites and the collinearity of these variables at Sampela 1. Nevertheless, these run-off associated stressors do constitute a genuine threat to many reefs in Indonesia (Edinger *et al.*, 1998; Edinger & Risk, 2013).

Coral mortality appeared to promote dead substrate availability, with an inverse correlation between the cover of these respective substrate types. Elsewhere, coral cover declines have been causally linked with increases in macroalgal cover (Hughes *et al.*, 1999; McManus & Polsenberg, 2004), but in

the Wakatobi macroalgae is uncommon (J. Marlow, personal observations). Wakatobi reefs appear to be neither dominated by coral nor macroalgae, instead the majority of the reef area is comprised of dead substrate (33%) and other live benthos (30%), as is found across much of the Indo-Pacific (Bruno *et al.*, 2009).

### Implications for reef management

Bioeroding sponges in the Wakatobi occupied 8.9% of the available dead reef substrate, which is somewhat lower than found on other reef systems; e.g. 20–81% in the Mexican Pacific and 4–30% of dead substrate on the GBR (Schönberg & Ortiz, 2009; Nava & Carballo, 2013). The high abundances on these reefs and other bioeroding sponge increases on degraded coral reefs are primarily attributed to the sponges' ability to exploit newly available calcareous habitat released by coral mortality (e.g. Lopez-Victoria & Zea, 2005; Schönberg & Ortiz, 2009; Schönberg *et al.*, 2017a). In the absence of baseline data, the spatial patterns of bioeroding sponge assemblages in this study could not be conclusively associated with coral mortality and prevented the recognition of a temporal sequence of events related to reef degradation. However, our findings of a negative correlation between coral cover and dead substrate, and the importance of the availability of this dead substrate for bioeroding sponges, supports our hypothesis that coral mortality can promote the abundance of bioeroding sponges in the region. Nevertheless the importance of dead substrate availability differed between species and had no overall association with sponge diversity. Importantly, most of the local species had an apparent low resilience to excessive sediment deposition and turbidity. This suggests that on reefs degraded by terrestrial run-off, some bioeroding sponge species may be prevented from exploiting increases in substrate availability. In these environments, the species assemblage is likely to be dominated by fewer resilient species. This has already been observed in the eastern Pacific, where Nava & Carballo (2013) found that although habitat availability promoted bioeroding sponge abundance, resilience to scouring by moving sediment was an important determinant of assemblage composition. In conclusion, not all factors that negatively affect corals are of benefit to bioeroding sponges. It is therefore vital to obtain a better understanding of the biological traits of bioeroding organisms, and of how they sustain or reduce the ecosystem health when reef function is stressed or degraded.

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### REFERENCES

- Alvarez-Filip L, Gill JA, Dulvy N.K., Perry A.L., Watkinson A.R. and Côté L.M. (2011) Drivers of region-wide declines in architectural complexity on Caribbean reefs. *Coral Reefs* 30, 1051–1060.
- Anderson M., Gorley R. and Clarke K. (2008) *PERMANOVA for PRIMER: guide to software and statistical methods*. Plymouth: PRIMER-E.
- Bannister R., Battershill C. and de Nys R. (2012) Suspended sediment grain size and mineralogy across the continental shelf of the Great Barrier Reef: impacts on the physiology of a coral reef sponge. *Continental Shelf Research* 32, 86–95.
- Bautista-Guerrero E., Carballo J.L., Aguilar-Camacho J.M. and Sifuentes-Romero I. (2016) Molecular and morphological differentiation of sympatric larvae of coral excavating sponges of genus *Thoosa*. *Zoomorphology* 135, 159–165.
- Bautista-Guerrero E., Carballo J.L. and Maldonado M. (2010) Reproductive cycle of the coral-excavating sponge *Thoosa misalolli* (Clonidae) from Mexican Pacific coral reefs. *Invertebrate Biology* 129, 285–296.
- Bell J.J., Davy S.K., Jones T., Taylor M.W. and Webster N.S. (2013) Could some coral reefs become sponge reefs as our climate changes? *Global Change Biology* 19, 2613–2624.
- Bell J.J., McGrath E., Biggerstaff A., Bates T., Bennett H., Marlow J. and Shaffer M. (2015) Sediment impacts on marine sponges. *Marine Pollution Bulletin* 94, 5–13.
- Bell J.J. and Smith D. (2004) Ecology of sponge assemblages (Porifera) in the Wakatobi region, south-east Sulawesi, Indonesia: richness and abundance. *Journal of the Marine Biological Association of the United Kingdom* 84, 581–591.
- Bell J.J. and Turner J.R. (2000) Factors influencing the density and morphometrics of the cup coral *Caryophyllia smithii* in Lough Hyne. *Journal of the Marine Biological Association of the United Kingdom* 80, 437–441.
- Bergman K.M. (1983) *Distribution and ecological significance of the boring sponge Cliona viridis on the Great Barrier Reef, Australia*. MSc thesis, McMaster University, Hamilton, Ontario, Canada.
- Biggerstaff A., Smith D.J., Jompa J. and Bell J.J. (2015) Photoacclimation supports environmental tolerance of a sponge to turbid low-light conditions. *Coral Reefs* 34, 1049–1061.
- Biggerstaff A., Smith D.J., Jompa J. and Bell J.J. (2017) Metabolic responses of a phototrophic sponge to sedimentation supports transitions to sponge-dominated reefs. *Scientific Reports* 7, 2725.
- Bruno J.F. and Selig E.R. (2007) Regional decline of coral cover in the Indo-Pacific: timing, extent, and subregional comparisons. *PLoS ONE* 2, e711.
- Bruno J.F., Sweatman H., Precht W.F., Selig E.R. and Schutte V.G. (2009) Assessing evidence of phase shifts from coral to macroalgal dominance on coral reefs. *Ecology* 90, 1478–1484.
- Burke L.M., Reyter K., Spalding M. and Perry A. (2011) *Reefs at risk revisited*. Washington, DC: World Resources Institute.
- Burnham K.P. and Anderson D.R. (2004) Multimodel inference: understanding AIC and BIC in model selection. *Sociological Methods and Research* 33, 261–304.
- Büttner E. and Siebler F. (2013) *The impact of simulated dredging on sponges from the East Australian coastal line*. Habilitation thesis, University of Stuttgart, Germany.
- Calcinaï B., Azzini F., Bavestrello G., Cerrano G., Pansini M. and Thung D.C. (2006) Boring sponges from the Ha Long Bay, Tonkin Gulf, Vietnam. *Zoological Studies* 45, 201–212.

- Cacinaï B., Bavestrello G. and Cerrano C.** (2005) Excavating sponge species from the Indo-Pacific Ocean. *Zoological Studies* 44, 5–18.
- Cacinaï B., Bavestrello G., Cerrano C. and Gaggero L.** (2008) Substratum microtexture affects the boring pattern of *Cliona albimarginata* (Clonidae, Demospongiae). In Wisshak M. and Tapanila L. (eds) *Current developments in bioerosion*. Berlin: Springer, pp. 203–211.
- Callahan M.K.** (2005) *Distribution of chonid sponges in the Florida Keys National Marine Sanctuary (FKNMS), 2001–2003*. MSc thesis. University of South Florida, Tampa, USA.
- Carballo J.L., Bautista-Guerrero E. and Leyte-Morales G.E.** (2008) Boring sponges and the modelling of coral reefs in the east Pacific Ocean. *Marine Ecology Progress Series* 356, 113–122.
- Carballo J.L., Bautista E., Nava H., Cruz-Barraza J.A. and Chávez J.A.** (2013) Boring sponges, an increasing threat for coral reefs affected by bleaching events. *Ecology and Evolution* 3, 872–886.
- Carballo J.L., Sanchez-Moyano J.E. and Garcia-Gómez J.C.** (1994) Taxonomic and ecological remarks on boring sponges (Clonidae) from the straits of Gibraltar (southern Spain): tentative bioindicators? *Zoological Journal of the Linnean Society* 112, 407–424.
- Chaves-Fonnegra A., Feldheim K.A., Secord J. and Lopez J.V.** (2015) Population structure and dispersal of the coral-excavating sponge *Cliona delitrix*. *Molecular Ecology* 24, 1447–1466.
- Chaves-Fonnegra A. and Zea S.** (2007) Observations on reef coral undermining by the Caribbean excavating sponge *Cliona delitrix* (Demospongiae, Hadromerida). In Custódio M.R., Lôbo-Hajdu G., Hajdu E. and Murky G. (eds) *Porifera research. Biodiversity, innovation and sustainability*. Rio de Janeiro: National Museum, pp. 247–254.
- Chaves-Fonnegra A., Zea S. and Gómez M.L.** (2007) Abundance of the excavating sponge *Cliona delitrix* in relation to sewage discharge at San Andrés Island, SW Caribbean, Colombia. *Boletín de Investigaciones Marinas y Costeras* 36, 63–78.
- Cheal A., MacNeil M.A., Cripps E., Emslie M., Jonker M., Schaffelke B. and Sweatman H.** (2010) Coral-macroalgal phase shifts or reef resilience: links with diversity and functional roles of herbivorous fishes on the Great Barrier Reef. *Coral Reefs* 29, 1005–1015.
- Clarke K.R. and Gorley R.N.** (2006) *PRIMER v6: user manual/tutorial*. Plymouth: PRIMER-E.
- Clarke K.R., Somerfield P.J. and Chapman M.G.** (2006) On resemblance measures for ecological studies, including taxonomic dissimilarities and a zero-adjusted Bray–Curtis coefficient for denuded assemblages. *Journal of Experimental Marine Biology and Ecology* 330, 55–80.
- Cooper T.F., Ridd P.V., Ulstrup K.E., Humphrey C., Slivkoff M. and Fabricius K.E.** (2008) Temporal dynamics in coral bioindicators for water quality on coastal coral reefs of the Great Barrier Reef. *Marine and Freshwater Research* 59, 703–716.
- Crabbe J.M. and Smith D.J.** (2002) Comparison of two reef sites in the Wakatobi Marine National Park (SE Sulawesi, Indonesia) using digital image analysis. *Coral Reefs* 21, 242–244.
- Crabbe J.M. and Smith D.J.** (2005) Sediment impacts on growth rates of *Acropora* and *Porites* corals from fringing reefs of Sulawesi, Indonesia. *Coral Reefs* 24, 437–441.
- Cullen L.C., Pretty J., Smith D. and Pilgrim S.E.** (2007) Links between local ecological knowledge and wealth in indigenous communities of Indonesia: implications for conservation of marine resources. *International Journal of Interdisciplinary Social Sciences* 2, 289–299.
- De'ath G. and Fabricius K.E.** (2010) Water quality as a regional driver of coral biodiversity and macroalgae on the Great Barrier Reef. *Ecological Applications* 20, 840–850.
- De'ath G., Fabricius K.E., Sweatman H. and Puotinen M.** (2012) The 27-year decline of coral cover on the Great Barrier Reef and its causes. *Proceedings of the National Academy of Sciences USA* 109, 17995–17999.
- Devlin M.J., Da Silva E.T., Petus C., Wenger A., Zeh D., Tracey D., Álvarez-Romero J.G. and Brodie J.** (2013) Combining in-situ water quality and remotely sensed data across spatial and temporal scales to measure variability in wet season chlorophyll-a: Great Barrier Reef lagoon (Queensland, Australia). *Ecological Processes* 2, 31.
- Doty M.S.** (1971) Measurement of water movement in reference to benthic algal growth. *Botanica Marina* 14, 32–35.
- Edinger E.N., Limmon G.V., Jompa J., Widjatmoko W., Heikoop J.M. and Risk M.J.** (1998) Reef degradation and coral biodiversity in Indonesia: effects of land-based pollution, destructive fishing practices and changes over time. *Marine Pollution Bulletin* 36, 617–630.
- Edinger E.N., Limmon G.V., Jompa J., Widjatmoko W., Heikoop J.M. and Risk M.J.** (2000) Normal coral growth rates on dying reefs: are coral growth rates good indicators of reef health? *Marine Pollution Bulletin* 40, 404–425.
- Edinger E.N. and Risk M.J.** (2013) Effect of land-based pollution on central Java coral reefs. *Journal of Coastal Development* 3, 593–613.
- English S.S., Wilkinson C.C. and Baker V.V.** (1994) *Survey manual for tropical marine resources. Line intercept transect*. Townsville: Australian Institute of Marine Science, pp. 34–51.
- Enochs I.C. and Manzello D.P.** (2012) Species richness of motile cryptofauna across a gradient of reef framework erosion. *Coral Reefs* 31, 653–661.
- Ertfemeijer P.L., Riegl B., Hoeksema B.W. and Todd P.A.** (2012) Environmental impacts of dredging and other sediment disturbances on corals: a review. *Marine Pollution Bulletin* 64, 1737–1765.
- Fabricius K.E.** (2005) Effects of terrestrial runoff on the ecology of corals and coral reefs: review and synthesis. *Marine Pollution Bulletin* 50, 125–146.
- Fabricius K.E.** (2011) Factors determining the resilience of coral reefs to eutrophication: a review and conceptual model. In Dubinsky Z. and Stambler N. (eds) *Coral reefs: an ecosystem in transition*. Dordrecht: Springer, pp. 493–505.
- Fang J.K.H., Schönberg C.H.L., Hoegh-Guldberg O. and Dove S.** (2017) Symbiotic plasticity of *Symbiodinium* in a common excavating sponge. *Marine Biology* 164, 104.
- Fang J.K.H., Schönberg C.H.L., Mello-Athayde M.A., Hoegh-Guldberg O. and Dove S.** (2014) Effects of ocean warming and acidification on the energy budget of an excavating sponge. *Global Change Biology* 20, 1043–1054.
- Glynn P.W.** (1997) Bioerosion and coral-reef growth: a dynamic balance. In Birkeland C. (ed.) *Life and death of coral reefs*. New York, NY: Springer, pp. 68–95.
- Golbuu Y., Van Woesik R., Richmond R.H., Harrison P. and Fabricius K.E.** (2011) River discharge reduces reef coral diversity in Palau. *Marine Pollution Bulletin* 62, 824–831.
- González-Rivero M., Yakob L. and Mumby P.J.** (2011) The role of sponge competition on coral reef alternative steady states. *Ecological Modelling* 222, 1847–1853.
- Graham N.A. and Nash K.L.** (2012) The importance of structural complexity in coral reef ecosystems. *Coral Reefs* 32, 315–326.
- Hennige S.J., Smith D.J., Perkins R., Consalvey M., Paterson D.M. and Suggett D.J.** (2008) Photoacclimation, growth and distribution of massive coral species in clear and turbid waters. *Marine Ecology Progress Series* 369, 77–88.

- Hennige S.J., Smith D.J., Walsh S.J., McGinley M.P., Warner M. E. and Suggett D.J. (2010) Acclimation and adaptation of scleractinian coral communities along environmental gradients within an Indonesian reef system. *Journal of Experimental Marine Biology and Ecology* 391, 143–152.
- Holmes K.E., Edinger E.N., Limmon G.V. and Risk M.J. (2000) Bioerosion of live massive corals and branching coral rubble on Indonesian coral reefs. *Marine Pollution Bulletin* 40, 606–617.
- Holmes G., Ortiz J.C. and Schönberg C.H.L. (2009) Bioerosion rates of the sponge *Cliona orientalis* Thiele, 1900: spatial variation over short distances. *Facies* 55, 203–311.
- Hughes T., Szmant A.M., Steneck R., Carpenter R. and Miller S. (1999) Algal blooms on coral reefs: what are the causes? *Limnology and Oceanography* 44, 1583–1586.
- Jokiel P.L. and Morrissey J.I. (1993) Water motion on coral reefs – evaluation of the clod card technique. *Marine Ecology Progress Series* 93, 175–181.
- Kennedy E.V., Perry C.T., Halloran P.R., Fine M., Carricart-Ganivet J.P., Iglesias-Prieto R., Form A., Wisshak M., Schönberg C.H.L. and Mumby P.J. (2013) Avoiding coral structural collapse requires local and global action. *Current Biology* 23, 912–918.
- Lapointe B.E., Barile P.J. and Matzie W.R. (2004) Anthropogenic nutrient enrichment of seagrass and coral reef communities in the Lower Florida Keys: discrimination of local versus regional nitrogen sources. *Journal of Experimental Marine Biology and Ecology* 308, 23–58.
- Lindgren N.G. (1897) Beitrag zur Kenntniss der Spongienfauna des Malaischen Archipels und der Chinesischen Meere. *Zoologischer Anzeiger* 547, 480–487.
- López-Victoria M. and Zea S. (2004) Storm-mediated coral colonization by an excavating Caribbean sponge. *Climate Research* 26, 251–256.
- López-Victoria M. and Zea S. (2005) Current trends of space occupation by encrusting excavating sponges on Colombian coral reefs. *Marine Ecology* 26, 33–41.
- Mallela J. and Perry C. (2007) Calcium carbonate budgets for two coral reefs affected by different terrestrial runoff regimes, Rio Bueno, Jamaica. *Coral Reefs* 26, 129–145.
- Mariani S., Uriz M.J. and Turon X. (2000) Larval bloom of the oviparous sponge *Cliona viridis*: coupling of larval abundance and adult distribution. *Marine Biology* 137, 783–790.
- Marlow J., Smith D., Werorilang S. and Bell J. (in press) Sedimentation limits the erosion rate of a bioeroding sponge. *Marine Ecology* e12483 doi: 10.1111/mec.12483.
- McManus J.W. and Polsenberg J.F. (2004) Coral-algal phase shifts on coral reefs: ecological and environmental aspects. *Progress in Oceanography* 60, 263–279.
- McMellor S. and Smith D.J. (2010) Coral reefs of the Wakatobi: abundance and biodiversity. In Clifton I., Unsworth R.K.F. and Smith D.J. (eds) *Marine research and conservation in the Coral Triangle: the Wakatobi National Park*. New York, NY: Nova Publishers, pp. 11–26.
- Mueller B., de Goeij J.M., Vermeij M.J., Mulders Y., van der Ent E., Ribes M. and van Duyl F.C. (2014) Natural diet of coral-excavating sponges consists mainly of dissolved organic carbon (DOC). *PLoS ONE* 9, e90152.
- Mumby P.J. (2009) Phase shifts and the stability of macroalgal communities on Caribbean coral reefs. *Coral Reefs* 28, 761–773.
- Muricy G. (1991) Structure des peuplements de spongiaires autour de l'égot de Cortiou (Marseille, France). *Vie et Milieu* 41, 205–221.
- Nava H. and Carballo J.L. (2008) Chemical and mechanical bioerosion of boring sponges from Mexican Pacific coral reefs. *Journal of Experimental Biology* 211, 2827–2831.
- Nava H. and Carballo J.L. (2013) Environmental factors shaping boring sponge assemblages at Mexican Pacific coral reefs. *Marine Ecology* 34, 269–279.
- Nava H., Ramirez-Herrera M.T., Figueroa-Camacho A.G. and Villegas-Sanchez B.M. (2014) Habitat characteristics and environmental factors related to boring sponge assemblages on coral reefs near populated coastal areas on the Mexican eastern Pacific coast. *Marine Biodiversity* 44, 45–54.
- Norström A.V., Nyström M., Lokrantz J. and Folke C. (2009) Alternative states on coral reefs: beyond coral-macroalgal phase shifts. *Marine Ecology Progress Series* 376, 295–306.
- Ogston A.S., Storlazzi C.D., Field M.E. and Presto M.K. (2004) Sediment resuspension and transport patterns on a fringing reef flat, Molokai, Hawaii. *Coral Reefs* 23, 559–569.
- Pang R.K. (1973) The ecology of some Jamaican excavating sponges. *Bulletin of Marine Science* 23, 227–243.
- Perry C.T., Spencer T. and Kench P. (2008) Carbonate budgets and reef production states: a geomorphic perspective on the ecological phase-shift concept. *Coral Reefs* 27, 853–866.
- Pineda M.C., Strehlow B., Duckworth A., Doyle J., Jones R. and Webster N.S. (2016) Effects of light attenuation on the sponge holobiont – implications for dredging management. *Scientific Reports* 6, 39038.
- Pineda M.C., Strehlow B., Kamp J., Duckworth A., Jones R. and Webster N.S. (2017a) Effects of combined dredging-related stressors on sponges: a laboratory approach using realistic scenarios. *Scientific Reports* 7, 5155.
- Pineda M.C., Strehlow B., Sternal M., Duckworth A., Haan J.D., Jones R. and Webster N.S. (2017b) Effects of sediment smothering on the sponge holobiont with implications for dredging management. *Scientific Reports* 7, 5156.
- Pineda M.C., Strehlow B., Sternal M., Duckworth A., Jones R. and Webster N.S. (2017c) Effects of suspended sediments on the sponge holobiont with implications for dredging management. *Scientific Reports* 7, 4925.
- Pollock F.J., Lamb J.B., Field S.N., Heron S.F., Schaffelke B., Shedrawi G., Bourne D.G. and Willis B.L. (2014) Sediment and turbidity associated with offshore dredging increase coral disease prevalence on nearby reefs. *PLoS ONE* 9, e102498.
- Powell A., Smith D.J., Hepburn L.J., Jones T., Berman J., Jompa J. and Bell J.J. (2014) Reduced diversity and high sponge abundance on a sedimented Indo-Pacific reef system: implications for future changes in environmental quality. *PLoS ONE* 9, e85253.
- Ramsby B.D., Hoogenboom M.O., Whalan S., Webster N.S. and Thompson A. (2017) A decadal analysis of bioeroding sponge cover on the inshore Great Barrier Reef. *Scientific Reports* 7, 2706.
- Reis M.A.C. and Leão Z.M.A.N. (2000) Bioerosion rate of the sponge *Cliona celata* (Grant 1826) from reefs in turbid waters, north Bahia, Brazil. In Moosa M.K., Soemodihardjo S., Soegiarto A., Romimohtarto K., Nontji A., Soekarno and Suharsono (eds) *Proceedings of the 9th International Coral Reef Symposium*, Bali, Indonesia 23–27 October 2000. Volume 1, pp. 273–278.
- Reiswig H.M. (1971) Particle feeding in natural populations of three marine demosponges. *Biological Bulletin* 141, 568–591.
- Risk M.J., Sammarco P.W. and Edinger E.N. (1995) Bioerosion in *Acropora* across the continental shelf of the Great Barrier Reef *Coral Reefs* 14, 79–86.

- Rose C.S. and Risk M.J. (1985) Increase in *Cliona delitrix* infestation of *Montastrea cavernosa* heads on an organically polluted portion of the Grand Cayman fringing reef. *Marine Ecology* 6, 345–363.
- Rosell D. and Uriz M.J. (1991) *Cliona viridis* (Schmidt, 1862) and *Cliona nigricans* (Schmidt, 1862) (Porifera, Hadromerida): evidence which shows they are the same species. *Ophelia* 33, 45–53.
- Rützler K. (2002) Impact of crustose clonid sponges on Caribbean reef corals. *Acta Geologica Hispanica* 37, 61–72.
- Schönberg C.H.L. (2000) Bioeroding sponges common to the central Australian Great Barrier Reef: descriptions of three new species, two new records, and additions to two previously described species. *Senckenbergiana Marina* 30, 161–221.
- Schönberg C.H.L. (2001) Small-scale distribution of Great Barrier Reef bioeroding sponges in shallow water. *Ophelia* 55, 39–54.
- Schönberg C.H.L. (2002) Substrate effects on the bioeroding demosponge *Cliona orientalis*. 1. Bioerosion rates. *Marine Ecology* 23, 313–326.
- Schönberg C.H.L. (2008) A history of sponge erosion: from past myths and hypotheses to recent approaches. In Wisshak M. and Tapanila L. (eds) *Current developments in bioerosion*. Berlin: Springer, pp. 165–202.
- Schönberg C.H.L. (2015a) Monitoring bioeroding sponges: using rubble, quadrat, or intercept surveys? *Biological Bulletin* 228, 137–155.
- Schönberg C.H.L. (2015b) Self-cleaning surfaces in sponges. *Marine Biodiversity* 45, 623–624.
- Schönberg C.H.L. (2016a) Happy relationships of marine sponges with sediments – a review and some observations from Australia. *Journal of the Marine Biological Association of the United Kingdom* 96, 493–514.
- Schönberg C.H.L. (2016b) *Project 6.1 – effects of dredging on filter feeder communities, with a focus on sponges. Final report of project 6.1 of the Dredging Science Node of the Western Australian Marine Science Institution*. Perth: Australian Institute of Marine Science, 127 pp.
- Schönberg C.H.L., Fang J.K.H. and Carballo J.L. (2017a) Bioeroding sponges and the future of coral reefs. In Bell J.J. and Carballo J.L. (eds) *Climate change, ocean acidification and sponges*. Dordrecht: Springer.
- Schönberg C.H.L., Fang J.K.H., Carreiro-Silva M., Tribollet A. and Wisshak M. (2017b) Bioerosion: the other ocean acidification problem. *ICES Journal of Marine Science* 74, 895–925.
- Schönberg C.H.L. and Ortiz J. (2009) Is sponge bioerosion increasing? *Proceedings of the 11th International Coral Reef Symposium*, Fort Lauderdale, Florida, 7–11 July 2008, Volume 1, pp. 520–523.
- Soekarno R. (1989) Comparative studies on the status of Indonesian coral reefs. *Netherlands Journal of Sea Research* 23, 215–222.
- Tajuddah M., Mustafá A., Pangerang U.K. and Yasidi F. (2012) Application of satellite multi-sensor techniques to detect upwellings and potential fishing grounds in Wakatobi Marine National Park, Southeast Sulawesi, Indonesia. *Aquatic Ecosystem Health & Management* 15, 303–310.
- Tribollet A., Decherf G., Hutchings P. and Peyrot-Clausade M. (2002) Large-scale spatial variability in bioerosion of experimental coral substrates on the Great Barrier Reef (Australia): importance of microborers. *Coral Reefs* 21, 424–432.
- Tribollet A. and Golubic S. (2005) Cross-shelf differences in the pattern and pace of bioerosion of experimental carbonate substrates exposed for 3 years on the northern Great Barrier Reef, Australia. *Coral Reefs* 24, 422–434.
- Wisshak M. and Tapanila L. (2008) *Current developments in bioerosion*. Berlin: Springer.
- and
- Wyrski K. (1961) *Physical oceanography of the Southeast Asian Waters. NAGA report volume 2*. San Diego, CA: Scripps Institution of Oceanography, 195 pp.

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## APPENDIX

**Table A1.** Results of bivariate correlations (Spearman's Rank,  $r_s$ ) comparing the standardized abundance of individual species of bioeroding sponges with environmental factors. Only the three most dominant species are shown.

| Factor                    | Species                  | Samples included |                  | Samples excluded |                  |
|---------------------------|--------------------------|------------------|------------------|------------------|------------------|
|                           |                          | $r_s$            | $P$              | $r_s$            | $P$              |
| Depth of settled sediment | <b>C. cf. aurivillii</b> | <b>-0.248</b>    | <b>0.023</b>     | <b>-0.343</b>    | <b>0.003</b>     |
|                           | C. aff. viridis sp. B    | -0.078           | 0.480            | 0.125            | 0.296            |
|                           | <b>S. cf. vagabunda</b>  | <b>0.260</b>     | <b>0.017</b>     | <b>0.601</b>     | <b>&lt;0.001</b> |
| Water movement            | C. cf. aurivillii        | 0.062            | 0.576            | 0.034            | 0.779            |
|                           | C. aff. viridis sp. B    | -0.148           | 0.179            | -0.139           | 0.244            |
|                           | <b>S. cf. vagabunda</b>  | <b>-0.438</b>    | <b>&lt;0.001</b> | <b>-0.471</b>    | <b>&lt;0.001</b> |
| Turbidity                 | <b>C. cf. aurivillii</b> | -                | -                | 0.089            | 0.455            |
|                           | C. aff. viridis sp. B    | -                | -                | 0.177            | 0.138            |
|                           | <b>S. cf. vagabunda</b>  | -                | -                | <b>0.472</b>     | <b>&lt;0.001</b> |
| Chl- <i>a</i>             | C. cf. aurivillii        | -0.125           | 0.258            | -                | -                |
|                           | C. aff. viridis sp. B    | -0.027           | 0.810            | -                | -                |
|                           | <b>S. cf. vagabunda</b>  | <b>-0.031</b>    | <b>0.776</b>     | -                | -                |

Significant correlations are displayed in bold

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