

Yusuf_2019_IOP_Conf._Ser._3A
_Earth_Environ._Sci._253_0120
22.pdf
by

FILE	YUSUF_2019_IOP_CONF._SER._3A_EARTH_ENVIRON._SCI._253_012022 .PDF (777.86K)		
TIME SUBMITTED	01-OCT-2020 07:04AM (UTC+0700)	WORD COUNT	3883
SUBMISSION ID	1401785863	CHARACTER COUNT	20128

PAPER • OPEN ACCESS

Effect of increased CO₂ concentration on the growth rate of *Isopora palifera* and *Acropora hyacinthus* from different cross-shelf reef zones

4
To cite this article: M Y Yusuf *et al* 2019 *IOP Conf. Ser.: Earth Environ. Sci.* **253** 012022

View the [article online](#) for updates and enhancements.

Effect of increased CO₂ concentration on the growth rate of *Isopora palifera* and *Acropora hyacinthus* from different cross-shelf reef zones

M Y Yusuf, N Rukminasari, D Yanuarita, J Jompa and Suharto

Faculty of Fisheries and Marine Science, Hasanuddin University, Makassar, Indonesia

Email: yusfiyusuf@outlook.com

Abstract. The rise in atmospheric carbon dioxide (CO₂) concentration due to emissions associated with economic development is altering ocean chemistry, a process known as ocean acidification. The resultant decrease in oceanic pH will affect marine organisms, in particular those which build their skeletons through calcification such as scleractinian corals. The aim of this research was to analyse the likely impact of changing CO₂ concentrations and thus seawater pH on the growth rate of two common corals, *Isopora palifera* and *Acropora hyacinthus*. This research was conducted at the Marine, Coastal and Small Island Research Centre, Universitas Hasanuddin, Indonesia. Samples of *Isopora palifera* and *Acropora hyacinthus* were collected along an inshore-offshore cross-shelf gradient from 3 sites: Pulau Karanrang (inner zone), Pulau Badi (intermediate zone), and Pulau Kapoposang (outer zone). A fully randomised research design was used with three replicates for each of three CO₂ treatments: 390 ppm (control), 550 ppm (2030 prediction), 1000 ppm (2050 prediction). The samples were weighed weekly for 1 month (digital balance, accuracy 0.1 mg). ANOVA analysis with post hoc Tukey Test showed a significant ($p < 0.05$) between treatment difference in growth rate for both *Isopora palifera* and *Acropora hyacinthus* ($P < 0.05$). The corals from all three zones exhibited positive growth at 390 ppm CO₂, and negative growth at CO₂ concentrations of 550 ppm and 1000 ppm.

1. Introduction

The phenomenon of climate change is currently a popular topic for research around the world. Increased human economic activities derived from the use of fossil fuels, such as oil and coal, have an impact on increasing greenhouse gas emissions, in particular carbon dioxide (CO₂). These emissions are causing an increase in atmospheric CO₂ concentrations. The scientific consensus estimate is that the average carbon dioxide partial pressure concentration (pCO₂) in the atmosphere has increased rapidly from around 280 ppm in pre-industrial times to 390 ppm in 2005 [1]. It has been predicted that by 2030 pCO₂ concentrations would likely reach 450 ppm [2], and are predicted to reach 900 ppm by the end of the 21st century [3].

In the natural carbon cycle, atmospheric carbon dioxide gas levels are determined by the balance of reactions in the atmosphere, land and ocean. Much of the CO₂ gas in the atmosphere is absorbed by plants on land and some is dissolved into the ocean and utilized by marine organisms [1]. Atmospheric carbon dioxide (CO₂) absorbed by the oceans reacts with sea water (H₂O) to form carbonic acid (H₂CO₃) which dissolves rapidly to form ionised hydrogen (H⁺) ions (an acid) and

carbonate, HCO_3^- (a base) [4]. The increasing abundance of hydrogen ions in the ocean reduces the pH of seawater, a phenomenon known as ocean acidification [5,6,7].

The results of previous research [8,9,10] show that acidification of sea water due to increased carbon dioxide concentration can affect the survival of marine organisms which calcify during some or all life cycle stages, including calcareous macroalgae, shellfish, and coral reefs. In calcareous macroalgae, decreasing water pH levels can interfere with the calcification process of *Halimeda* sp.[8]. Calcite formation in shellfish can become harder due to ocean acidification, and can interfere with the biomineralisation process [9]. Ocean acidification can also reduce the availability of carbonate needed by coral animals to form calcium carbonate skeletons, so that growth can be inhibited [2,10,11,12].

The Spermonde Islands in the southern Makassar Strait, off the southwest coast of Sulawesi Island, have extensive coral reefs, influenced by the Makassar Strait water mass [13]. The Spermonde Islands can be divided into four zones, starting from the inner zone to the outer zone [14]. Coral biodiversity is fairly high (78 genera with a total of 262 species recorded), of which around 80-87% are in the outer reef areas. However, in 1996 [15] it was noted that, compared to data recorded in the early 1980's some of the same locations [14], live coral cover and diversity had been reduced by around 20% within 12 years. This appeared to be the result of high levels of sedimentation and eutrophication originating from anthropogenic activities.

Although an increase in the concentration of carbon dioxide in the atmosphere has been detected world-wide, there was a lack of scientific information on the impact of increasing carbon dioxide concentrations on coral condition in the Spermonde Islands. It was therefore considered important to evaluate the impact of increased carbon dioxide concentrations on the genus *Acropora*, which has been used as a reference for coral health in different zones within the Spermonde waters.

The purpose of this study was to examine the effect of increasing carbon dioxide, at different concentrations, on the growth rate of *Isopora palifera* and *Acropora hyacinthus*. The research was planned on a laboratory scale, and compared the growth rates of *Isopora palifera* and *Acropora hyacinthus* from each Spermonde zone under different carbon dioxide concentrations.

2. Materials and methods

2.1. Time and place

This study was carried out in April - May 2018. *Acropora* coral samples were collected from sampling stations on several islands across the Spermonde Archipelago (Kang, Badi, and Kapoposang Islands). The experimental research was conducted at the Research Center for Marine, Coastal and Small Islands, Hasanuddin University, Makassar.

2.2. Setting up the experimental CO₂ system

The CO₂ system is a tool that was assembled from a number of components, including supporting equipment such as a CO₂ supply, an O₂ compressor and a mass flow controller (MFC) (Figure 1). This system functions as a regulator of carbon dioxide (CO₂) concentration in the water. Carbon dioxide gas from CO₂ gas cylinders and oxygen from the O₂ compressors both enter the mass flow controller. The MFC regulates the flow rate and has CO₂ meters with digital displays so that the rate of carbon dioxide concentration flowing into the aquaria can be measured and regulated. For this research, the mass flow controller was set for two concentrations: (i) 550 ppm pCO₂ (CO₂ gas flow rate range 7.95 - 8 mL/min and O₂ range 2:49 to 2:55 L/min) and (ii) 1000 ppm pCO₂ (CO₂ gas flow rate range 9.95 - 10 mL/min and O₂ range 1 - 1:10 L/min)..



Figure 1. CO₂ system components:
 (a) CO₂ gas cylinders
 (b) O₂ compressor
 (c) Mass flow controller
 (d) CO₂ meter

2.3. Research Procedures

2.3.1. Collection and acclimatization of coral samples. Coral sampling sites in the Spermonde Islands were selected to represent the cross-shelf zones. Karanrang Island represents the inner zone, Badi Island represents the middle zone, and Kapoposang Island the outer zone. Stations at each sampling site were selected at a depth of 3 - 5 meters, at which *Acropora* colonies are typically most abundant. The weight of each sample (fragment) collected from *Acropora hyacinthus* and *Isopora palifera* coral colonies was <100 grams. The fragments were taken using pliers, then transplanted (using glue) on a mica substrate and placed at the same depth as the 5-month sampling location, so that the transplanted samples (fragments) could adjust (recover) before being used in the experiment.

2.3.2. Experimental Design. This research used a factorial design with 5 factors: CO₂ concentration treatment, sampling zone, species, fragments, and replicates. There were three pCO₂ concentration levels (390 ppm, 550 ppm, and 1000 ppm), each with three replicates (Figure 2).

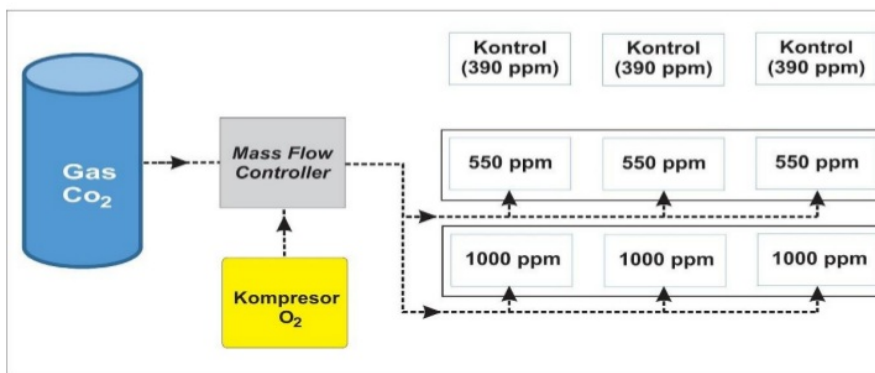


Figure 2. Research Layout

The 390 ppm concentration is the current ambient pCO₂ concentration [16], while 550 ppm and 1000 ppm are the predicted pCO₂ concentration in 2030 and 2100 [1,2,3]. This concentration

difference assumes that an increase in CO₂ results in a decrease in seawater pH [5,6,7]. The three sampling zones were: inner zone (Karanrang Island), middle zone (Badi Island), and outer zone (Kapoposang Island). The two species were *Isopora palifera* and *Acropora hyacinthus*. The fragment factor comprised fragments A and B. Thus, the total number of experimental units (fragments) in this study design is $3 \times 3 \times 3 \times 2 \times 2 = 108$ coral fragments. During the study, water quality parameters were also recorded and water changed once a week. Temperature, salinity, dissolved oxygen, and pH were measured using water quality meters, alkalinity using a Mini Alkalinity Titrator.

2.3.3. Sample weight and growth rate. Test organisms (*Acropora* coral fragments) were placed into an aquarium (90 cm x 39 cm x 35 cm) for 1 month. Before being placed in the aquarium, each fragment was weighed to obtain the initial weight (t_0) using analytical digital scales with an accuracy of 0.1 mg (0.0001 g). The fragments were then weighed weekly, so that the rate of weight change could be calculated using the formula:

$$C = t_n - t_0$$

Where: C = weight change (mg); t_n = weight in week-n (mg); t_0 = initial weight (mg)

2.4. Data Analysis

The weight change data were analysed descriptively, tabulated and presented in graphic (histogram) form. Prior to conducting statistic tests, the data on growth rates of *Isopora palifera* and *Acropora hyacinthus* from different zones were first tested for normality. If the data were normally distributed, an Analysis of variance (Factorial ANOVA) was applied, otherwise the non-parametric Kurskall-Wallis test was applied. Data analysis was performed in SPSS v.18.

3. Results and Discussion

3.1. Results

3.1.1. *Acropora hyacinthus* weight change. The weight change of *Acropora hyacinthus* under each treatment (Figure 3) shows that under the 390 ppm pCO₂ treatment, *Acropora hyacinthus* from all three zones increased in weight.

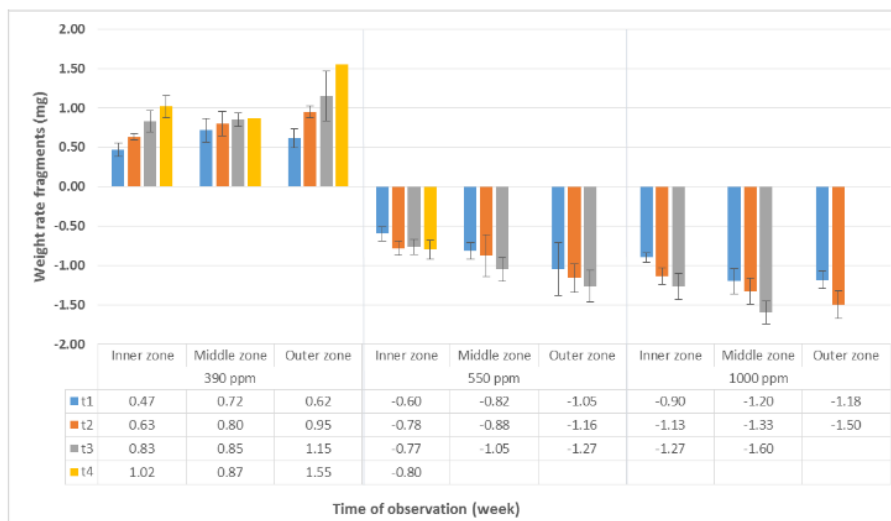


Figure 3. Weight change of *Acropora hyacinthus* fragments under the three pCO₂ treatments

At 550 ppm pCO₂, the coral fragments decreased in weight every week, with inner zone fragments having the lowest decrease and outer reef fragments the highest. Fragments from the middle and outer zones experienced bleaching in the third week. At 1000 ppm pCO₂ concentration the coral fragments from all coral reef distribution zones in Spermonde waters experienced a greater decrease in weight rate than under the 550 ppm treatment, and bleached more rapidly, after 2 to 3 weeks. Weight loss was higher and bleaching occurred more rapidly in outer than inner zone fragments.

A One-Way Anova with Tukey post hoc test showed the weight change of *Acropora hyacinthus* fragments was significantly different ($P < 0.05$) between the 390 ppm, 550 ppm, and 1000 ppm carbon dioxide concentration treatments. However the Kruskal Wallis test showed the weight changes were not significantly different ($P > 0.05$) between zones

3.1.2. *Isopora palifera* weight change. The weight changes *Isopora palifera* fragments (Figure 4) show that under the 390 ppm pCO₂ treatment *Isopora palifera* coral fragments from all three zones increased in weight during the experiment. At 550 ppm and 1000 ppm pCO₂, *Isopora palifera* from all zones in Spermonde waters experienced a decrease in weight every week, with a higher weight loss at 1000 ppm than 550 ppm.

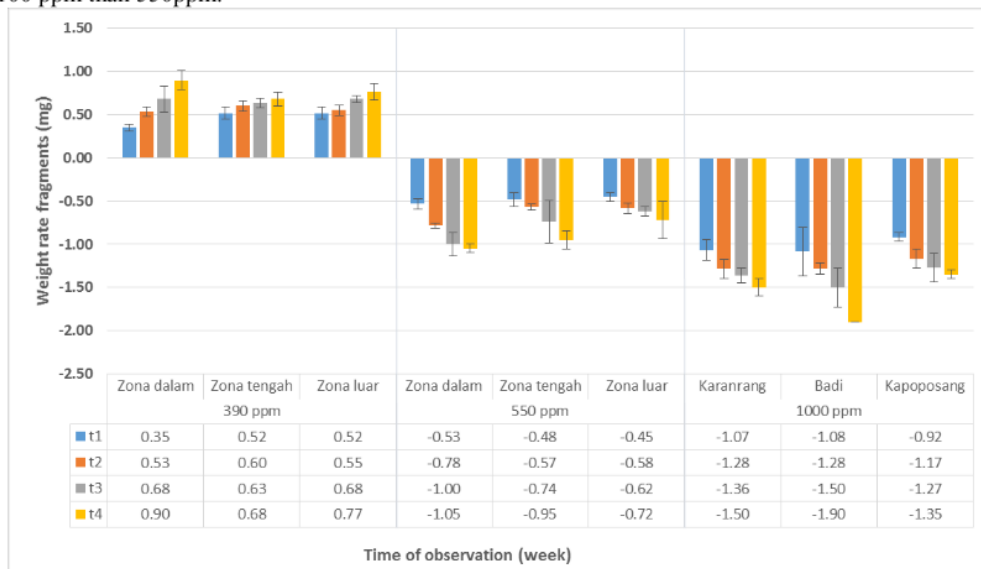


Figure 4. Weight change of *Isopora palifera* coral fragments under the three pCO₂ treatments

Based on the results of the One-Way Anova test with post hoc tests Tukey showed that the weight rate of coral fragments *Isopora palifera* between treatments of 390 ppm, 550 ppm, and 1000 ppm carbon dioxide concentration was significantly different ($P < 0.05$). While the weight rate of coral fragments *Acropora palifera* was not significantly different ($P > 0.05$) between inner zones, middle zones and outer zones.

3.1.3. Water quality parameters. The water quality parameters for each experimental unit (aquarium) (Table 1) show that the pH values and other parameters measured were similar between replicates within each treatment. Parameters which varied significantly between treatments were DO, Alkalinity, and CO₂ (aq).

Table 1. Water quality parameters by treatment and replicate

Parameter	Replicate	$p\text{CO}_2$		
		390 ppm	550 ppm	1000 ppm
pH	I	8.03 ± 0.05	7.91 ± 0.04	7.76 ± 0.05
	II	8.02 ± 0.07	7.91 ± 0.04	7.75 ± 0.06
	III	8.02 ± 0.07	7.93 ± 0.04	7.75 ± 0.07
DO (ppm)	I	6.71 ± 0.19	4.11 ± 0.32	3.03 ± 0.25
	II	7.04 ± 0.40	4.18 ± 0.21	3.17 ± 0.24
	III	6.80 ± 0.33	4.01 ± 0.24	3.33 ± 0.22
Salinity (ppt)	I	31.61 ± 0.53	32.28 ± 0.83	31.66 ± 0.65
	II	31.60 ± 0.60	32.03 ± 0.49	31.59 ± 0.61
	III	32.25 ± 0.85	31.96 ± 0.92	31.51 ± 0.96
Temperature(°C)	I	29.68 ± 0.69	29.69 ± 0.59	29.53 ± 0.74
	II	29.54 ± 0.62	29.09 ± 0.65	29.07 ± 0.72
	III	30.09 ± 0.78	29.99 ± 0.63	29.71 ± 0.82
Alkalinity (ppm)	I	92.98 ± 0.65	83.48 ± 0.90	76.84 ± 0.94
	II	93.18 ± 0.72	84.37 ± 0.54	75.95 ± 1.08
	III	93.04 ± 0.57	83.75 ± 0.23	75.04 ± 0.90
$\text{CO}_2(\text{aq})$ (ppm)	I	10.63 ± 0.34	16.15 ± 0.24	27.18 ± 0.15
	II	10.6 ± 0.32	16.28 ± 0.3	27.48 ± 0.25
	III	10.75 ± 0.54	16.43 ± 0.13	27.4 ± 0.22

3.2. Discussion

The results of this study show that under increased carbon dioxide (CO_2) concentrations there was a decrease in the coral growth rate, and indeed negative growth, in both *Acropora hyacinthus* and *Isopora palifera* (Figure 3 and Figure 4). The 390 ppm (control/ambient) CO_2 concentration treatment showed positive coral growth throughout the observation time; furthermore, growth rates varied with fragment size. Coral growth can be influenced by many factors including the age, shape and size of fragments or colonies [17]. Coral growth is also influenced by the reciprocal symbiotic relationship between polyps and zooxanthellae (*Symbiodinium* sp.), where zooxanthellae produce the oxygen and nutrients needed by polyps and polyps provide the zooxanthellae with living space and carbon dioxide for photosynthesis. The more polyps, the more zooxanthellae are found in the coral, so the calcification process should also be faster, resulting in increased coral growth rates. This hypothesis is supported by research showing that coral growth rate is influenced by the number of polyps [18]; furthermore, two and three polyps can use food more optimally compared to one polyp.

In the 550 and 1000 ppm $p\text{CO}_2$ treatments (Figure 3 and Figure 4), coral growth was negative. This indicates that under high CO_2 concentrations, the growth of *Acropora hyacinthus* and *Isopora palifera* corals is impeded. As the concentration of carbon dioxide dissolved in seawater increases, both pH and alkalinity decrease, and changes in the seawater carbonate balance reduce the level of aragonite, the mineral used by corals to build their limestone skeletons; these changes can disrupt the growth process [2,19].

Coral growth depends on environmental conditions, most of which are not fixed and tend to change due to disturbances originating from natural phenomena and human activities [20]. The water quality data (Table 1) show an increase in the partial carbon dioxide concentration ($p\text{CO}_2$) directly proportional to the concentration of carbon dioxide dissolved in water ($\text{CO}_2(\text{aq})$). This is in accordance with Henry's law which states that the amount of gas dissolved in a solution will be directly

proportional to the partial pressure of the gas in solution at equilibrium [21]. Research has shown [22,23,24] that at 400 ppm pCO₂, CO_{2(aq)} was 10.9 ppm, equivalent to pH 8.08; 597 ppm pCO₂ (pH = 7.89; CO_{2(aq)} = 16 ppm); 1004 ppm pCO₂ (pH = 7.7; CO_{2(aq)} = 27.4 ppm). Alkalinity reflects the buffering capacity of carbonate ions, with higher the alkalinity lowering fluctuations in pH [25]. Increased pCO₂ also causes reduced oxygen solubility in water, as seen in Table 1. Furthermore, the solubility of oxygen in water is influenced by temperature and salinity [26]; the higher the temperature and salinity, the lower the solubility of oxygen in water, and vice versa. Thus elevated temperatures and higher pCO₂ will act in synergy to reduce oxygen levels, which can negatively affect growth. Imbalances in the acid-base equilibrium can cause damage to exoskeleton components such as calcareous shells; the dissolving of calcium carbonate (CaCO₃) and accumulation of metabolic waste products can weaken an organism, disrupt oxygen transport processes, and if prolonged will cause death [27].

4. Conclusion

An increase in carbon dioxide (CO₂) concentration had a negative effect on the growth of *Acropora hyacinthus* and *Isopora palifera* in terms of weight. All corals originating from the inner zone (Karanrang Island), the middle zone (Badi Island) and the outer zone (Kapoposang Island) of the Spermonde Archipelago experienced an increase in weight at ambient partial concentration of carbon dioxide (390 ppm pCO₂). However the impact of predicted increases in pCO₂ under near future conditions is likely to inhibit coral weight gain and may result in high mortality or even extirpation of widespread and ecologically important corals.

2 Acknowledgments

This study was supported by a grant from USAID Peer Science Project Cycle 4, Coral Vulnerability Assessment to Temperature Stress (Bleaching) and Ocean Acidification in the Spermonde Archipelago: Conservation Strategies for Climate Resilience, COREMAP and the Ministry of Education and Culture Indonesia. The authors would like to thank the research team involved in Peer Science cycle 4: Abdul Wahid Hasdar, Abdul Rahman, Syaiful Bahri, Alinda Nurbaety Hasanah, Marwah Salam, Sudjriyana Mastari, Rosihan Anwar, Indra Adi Putra, Wulandari, Nur Rahmi, Rusman, and Fakhirah Ahmad who assisted the process of collecting the samples.

References

- [1] IPCC 2007 *Contribution of Working Groups I, II and III to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change* (Geneva: Intergovernmental Panel on Climate Change Synthesis Report)
- [2] Burke L, Reytar K, Spalding M and Perry A 2012 *Reefs at Risk, revisited* (Washington DC: World Resources Institute) 114 p
- [3] Houghton J T, Ding Y, Griggs D J, Noguer M *et al* 2001 *Climate Change 2001: The scientific Basis* (New York: Cambridge University Press)
- [4] OCB 2012 *Frequently Asked Question About Ocean Acidification* (UK Ocean Acidification Research Program: Ocean Carbon & Biogeochemistry) 25 pp
- [5] Doney S C, Fabry V J, Feely R A, Kleypas J A 2009 Ocean acidification: The other CO₂ problem *Ann Rev. Mar. Sci.* **1** 169-192
- [6] Nikinmaa M 2013 Climate change and ocean acidification – Interactions with aquatic toxicology *Aquat. Toxic.* **126** 365-372
- [7] Hendriks I E, Duarte C M, Olsen Y S, Steckbauer A *et al* 2014 Biological mechanisms supporting adaptation to ocean acidification in coastal ecosystems *Est. Coast. and Shelf Sci.* **30** 1-8
- [8] Rukminasari N, Nadiarti K and Awaluddin 2014 Pengaruh derajat keasaman (pH) air laut terhadap konsentrasi kalsium dan laju pertumbuhan *Halimeda* sp. *Torani* **24** 28-34
- [9] Fitzer S C, Vittert L, Bowman A, Kamenos N A *et al* 2015 Ocean acidification and temperature

- increase impact mussel shell shape and thickness: problematic for protection? *Ecol. Evol.* **5** 4875-4884
- [10] Feely R A, Sabine C L, Kee K, Barelson W *et al* 2004 Impact of anthropogenic CO₂ on the CaCO₃ system in the oceans *Science* **305** 362-366
- [11] Cao L, Caldeira K and Jain A K 2007 Effects of carbon dioxide and climate change on ocean acidification and carbonate mineral saturation *Geophys. Res. Lett.* **34** 5607
- [12] Silverman J, Lazar B, Cao L, Caldeira K and Erez J 2009 Coral reefs may start dissolving when atmospheric CO₂ doubles *Geophys. Res. Lett.* **36** 1-5
- [13] Rasyid J and Ibrahim A 2013 *Spermonde: Kondisi Oceanografi Versus Ikan Pelagis* (Makassar: Masagena Press) 124 pp
- [14] Moll H 1983 *Zonation and Diversity of Scleractinia on Reefs off S. W. Sulawesi* (Leiden: Thesis University Leiden)
- [15] Jompa J 1996 *Monitoring and Assessment of Coral Reefs on Spermonde Archipelago, South Sulawesi* (Canada: Thesis McMaster University)
- [16] Pajusalu L, Martin G, Pollumae A and Paalme T 2013 Results of laboratory and field experiments of the direct effect of increasing CO₂ on net primary production of macroalgal species in brackish - water ecosystems *Proc. Estonian Acad. Sci.* **62** 148-154
- [17] Jipriandi A, Pratomo H and Irawan 2013 Pertumbuhan karang *Acropora formosa* dengan teknik transplantasi pada ukuran fragmen yang berbeda *Jurnal Ilmiah Platax* **3** 90-100
- [18] Zulfikar and Soedharma D 2008 Teknologi fragmentasi buatan karang (*Caulastrea furcata* dan *Cynaria lacrimalis*) dalam upaya percepatan pertumbuhan pada kondisi terkontrol *Jurnal Natur Indonesia* **10** 76-82
- [19] Hoegh-Guldberg O, Hoegh-Guldberg H, Veron J E N 2009 *The Coral Triangle and Climate Change: Ecosystems, People and Societies at Risk* (Brisbane, Australia)
- [20] Castro P and Huber M E 2008 *Marine Biology (7th edition)* (McGraw-Hill 447 p.)
- [21] Vuong M D, Couvert A, Couriol C, Amrane A *et al* 2009 Determination of the Henry's Constants and the Mass Transfer Rate of VOCs in Solvent *Chem. Eng. J.* **150** 426-430
- [22] Cohen S, Krueger T and Fine M 2017 Measuring coral calcification under ocean acidification: methodological considerations for the 45Ca-uptake and total alkalinity anomaly technique *PeerJ* 1-21
- [23] Inoeu M, Suwa R, Suzuki A, Sakai K and Kawahata H 2011 Effects of seawater pH on growth and skeletal U/Ca ratios of *Acropora digitifera* coral polyps *Geophys. Res. Lett.* **38** 1-4
- [24] Iguichi A, Kumagai N H, Nakamura T, Suzuki A *et al* 2014 Responses of calcification of massive and encrusting corals to past, present, and near-future ocean carbon dioxide concentrations *Mar. Poll. Bull.* **89** 348-355
- [25] Yulfiperius, Toelihere M R, Affandi R, Sjafei D S 2004 Effect of Alkalinity on the Survival Rate and Growth of Lalawak Fish, *Barbodes* sp *Jurnal Iktiologi Indonesia* **4** 1-5
- [26] Colt J 1984 *Computation of Dissolved Gas Concentration in Water as Functions of Temperature, Salinity, and Pressure* (Amer. Fish. Soc. Spec. Pub. No. 14. 154 pp.)
- [27] Seibel B A and Walsh P J 2001 Potential impacts of CO₂ injection on deep-sea biota *Science* **294** 319-320

ORIGINALITY REPORT

% **13**
SIMILARITY INDEX

% **9**
INTERNET SOURCES

% **8**
PUBLICATIONS

% **4**
STUDENT PAPERS

PRIMARY SOURCES

1 DP Wijayanti, E Indrayanti, H Nuryadi, RA Dewi, A Sabdono. " Molecular Identification and Genetic Diversity of from Boo and Deer Island, Raja Ampat, West Papua ", IOP Conference Series: Earth and Environmental Science, 2018
Publication % **2**

2 journal.unhas.ac.id
Internet Source % **2**

3 eprints.unm.ac.id
Internet Source % **2**

4 Submitted to CSU, San Jose State University
Student Paper % **1**

5 www.medstarbloodless.org
Internet Source % **1**

6 "Complete issue pdf 73-3", ICES Journal of Marine Science, 2016
Publication % **1**

7 en.wikipedia.org
Internet Source % **1**

8	awsassets.panda.org Internet Source	% 1
9	www.freepatentsonline.com Internet Source	<% 1
10	pubs.iclarm.net Internet Source	<% 1
11	www.hindawi.com Internet Source	<% 1
12	nbn-resolving.de Internet Source	<% 1
13	www.yumpu.com Internet Source	<% 1
14	www.frontiersin.org Internet Source	<% 1
15	www.ppionline.org Internet Source	<% 1
16	willametteinitiative.org Internet Source	<% 1
17	Kucukali, S., S. Cokgor, and B. Kartal. "Temporal Variation of Dissolved Oxygen in a Mountain Stream Plunge Pool: An Example from Northern Turkey", World Environmental and Water Resource Congress 2006, 2006. Publication	<% 1

18

sites.nationalacademies.org

Internet Source

<% 1

19

onlinelibrary.wiley.com

Internet Source

<% 1

20

"Complete Issue pdf 74-4", ICES Journal of
Marine Science, 2017

Publication

<% 1

EXCLUDE QUOTES ON

EXCLUDE
BIBLIOGRAPHY ON

EXCLUDE MATCHES

< 5
WORDS